



## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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<b>(21) International Application Number:</b> PCT/US99/04608 <b>(22) International Filing Date:</b> 2 March 1999 (02.03.99) <b>(30) Priority Data:</b> 09/032,945                      2 March 1998 (02.03.98)                      US <b>(71) Applicant:</b> UNIVERSITY OF MASSACHUSETTS, A PUBLIC INSTITUTION OF HIGHER EDUCATION OF THE COMMONWEALTH OF MASSACHUSETTS, as represented by ITS AMHERST CAMPUS [US/US]; Office of Vice Chancellor for Research at Amherst, Amherst, MA 01002 (US). <b>(72) Inventors:</b> ROBL, James; 196 Old Enfield, Belchertown, MA 01007 (US). CIBELLI, Jose; 166 Village Park, Amherst, MA 01002 (US). STICE, Steven, L.; 468 Amherst Road, Belchertown, MA 01007 (US). <b>(74) Agent:</b> TESKIN, Robin, L.; Burns, Doane, Swecker & Mathis, L.L.P., P.O. Box 1404, Alexandria, VA 22313-1404 (US).		<b>(81) Designated States:</b> AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).  <b>Published</b> <i>With international search report.</i>
<b>(54) Title:</b> EMBRYONIC OR STEM-LIKE CELL LINES PRODUCED BY CROSS-SPECIES NUCLEAR TRANSPLANTATION <b>(57) Abstract</b> <p>An improved method of nuclear transfer involving the transplantation of differentiated donor cell nuclei into enucleated oocytes of a species different from the donor cell is provided. The resultant nuclear transfer units are useful for the production of isogenic embryonic stem cells, in particular human isogenic embryonic or stem cells. These embryonic or stem-like cells are useful for producing desired differentiated cells and for introduction, removal or modification, of desired genes, e.g., at specific sites of the genome of such cells by homologous recombination. These cells, which may contain a heterologous gene, are especially useful in cell transplantation therapies and for <i>in vitro</i> study of cell differentiation. Also, methods for improving nuclear transfer efficiency by genetically altering donor cells to inhibit apoptosis, select for a specific cell cycle and/or enhance embryonic growth and development are provided.</p>		

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1                   **EMBRYONIC OR STEM-LIKE CELL LINES PRODUCED BY**  
2                   **CROSS SPECIES NUCLEAR TRANSPLANTATION**

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6  
7                   **FIELD OF THE INVENTION**

8                   The present invention generally relates to the production of embryonic or  
9                   stem-like cells by transplantation of cell nuclei derived from animal or human  
10                  cells into enucleated animal oocytes of a species different from the donor nuclei.  
11                  The present invention more specifically relates to the production of primate or  
12                  human embryonic or stem-like cells by transplantation of the nucleus of a  
13                  primate or human cell into an enucleated animal oocyte, e.g., a primate or  
14                  ungulate oocyte and in a preferred embodiment a bovine enucleated oocyte.

15                 The present invention further relates to the use of the resultant embryonic  
16                 or stem-like cells, preferably primate or human embryonic or stem-like cells for  
17                 therapy, for diagnostic applications, for the production of differentiated cells  
18                 which may also be used for therapy or diagnosis, and for the production of  
19                 transgenic embryonic or transgenic differentiated cells, cell lines, tissues and  
20                 organs. Also, the embryonic or stem-like cells obtained according to the present  
21                 invention may themselves be used as nuclear donors in nuclear transplantation  
22                 or nuclear transfer methods for the production of chimeras or clons, preferably  
23                 transgenic cloned or chimeric animals.

24                 **BACKGROUND OF THE INVENTION**

25                 Methods for deriving embryonic stem (ES) cell lines *in vitro* from early  
26                 preimplantation mouse embryos are well known. (See, e.g., Evans et al., *Nature*,

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1 29:154-156 (1981); Martin, *Proc. Natl. Acad. Sci., USA*, 78:7634-7638 (1981)).  
2 ES cells can be passaged in an undifferentiated state, provided that a feeder layer  
3 of fibroblast cells (Evans et al., *Id.*) or a differentiation inhibiting source (Smith  
4 et al., *Dev. Biol.*, 121:1-9 (1987)) is present.

5 ES cells have been previously reported to possess numerous applications.  
6 For example, it has been reported that ES cells can be used as an *in vitro* model  
7 for differentiation, especially for the study of genes which are involved in the  
8 regulation of early development. Mouse ES cells can give rise to germline  
9 chimeras when introduced into preimplantation mouse embryos, thus demon-  
10 strating their pluripotency (Bradley et al., *Nature*, 309:255-256 (1984)).

11 In view of their ability to transfer their genome to the next generation, ES  
12 cells have potential utility for germline manipulation of livestock animals by  
13 using ES cells with or without a desired genetic modification. Moreover, in the  
14 case of livestock animals, e.g., ungulates, nuclei from like preimplantation  
15 livestock embryos support the development of enucleated oocytes to term (Smith  
16 et al., *Biol. Reprod.*, 40:1027-1035 (1989); and Keefer et al., *Biol. Reprod.*,  
17 50:935-939 (1994)). This is in contrast to nuclei from mouse embryos which  
18 beyond the eight-cell stage after transfer reportedly do not support the  
19 development of enucleated oocytes (Cheong et al, *Biol. Reprod.*, 48:958 (1993)).  
20 Therefore, ES cells from livestock animals are highly desirable because they may  
21 provide a potential source of totipotent donor nuclei, genetically manipulated or  
22 otherwise, for nuclear transfer procedures.

23 Some research groups have reported the isolation of purportedly  
24 pluripotent embryonic cell lines. For example, Notarianni et al., *J. Reprod. Fert.*  
25 *Suppl.*, 43:255-260 (1991), report the establishment of purportedly stable,  
26 pluripotent cell lines from pig and sheep blastocysts which exhibit some

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1 morphological and growth characteristics similar to that of cells in primary  
2 cultures of inner cell masses isolated immunosurgically from sheep blastocysts.  
3 (*Id.*) Also, Notarianni et al., *J. Reprod. Fert. Suppl.*, 41:51-56 (1990) discloses  
4 maintenance and differentiation in culture of putative pluripotential embryonic  
5 cell lines from pig blastocysts. Further, Gerfen et al., *Anim. Biotech*, 6(1):1-14  
6 (1995) disclose the isolation of embryonic cell lines from porcine blastocysts.  
7 These cells are stably maintained in mouse embryonic fibroblast feeder layers  
8 without the use of conditioned medium. These cells reportedly differentiate into  
9 several different cell types during culture (Gerfen et al., *Id.*).

10 Further, Saito et al., *Roux's Arch. Dev. Biol.*, 201:134-141 (1992) report  
11 bovine embryonic stem cell-like cell lines cultured which survived passages for  
12 three, but were lost after the fourth passage. Still further, Handyside et al.,  
13 *Roux's Arch. Dev. Biol.*, 196:185-190 (1987) disclose culturing of immunosurgi-  
14 cally isolated inner cell masses of sheep embryos under conditions which allow  
15 for the isolation of mouse ES cell lines derived from mouse ICMs. Handyside  
16 et al. (1987) (*Id.*), report that under such conditions, the sheep ICMs attach,  
17 spread, and develop areas of both ES cell-like and endoderm-like cells, but that  
18 after prolonged culture only endoderm-like cells are evident. (*Id.*)

19 Recently, Cherny et al., *Theriogenology*, 41:175 (1994) reported  
20 purportedly pluripotent bovine primordial germ cell-derived cell lines maintained  
21 in long-term culture. These cells, after approximately seven days in culture,  
22 produced ES-like colonies which stain positive for alkaline phosphatase (AP),  
23 exhibited the ability to form embryoid bodies, and spontaneously differentiated  
24 into at least two different cell types. These cells also reportedly expressed  
25 mRNA for the transcription factors OCT4, OCT6 and HES1, a pattern of  
26 homeobox genes which is believed to be expressed by ES cells exclusively.

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1           Also recently, Campbell et al., *Nature*, 380:64-68 (1996) reported the  
2           production of live lambs following nuclear transfer of cultured embryonic disc  
3           (ED) cells from day nine ovine embryos cultured under conditions which  
4           promote the isolation of ES cell lines in the mouse. The authors concluded based  
5           on their results that ED cells from day nine ovine embryos are totipotent by  
6           nuclear transfer and that totipotency is maintained in culture.

7           Van Stekelenburg-Hamers et al., *Mol. Reprod. Dev.*, 40:444-454 (1995),  
8           reported the isolation and characterization of purportedly permanent cell lines  
9           from inner cell mass cells of bovine blastocysts. The authors isolated and  
10          cultured ICMs from 8 or 9 day bovine blastocysts under different conditions to  
11          determine which feeder cells and culture media are most efficient in supporting  
12          the attachment and outgrowth of bovine ICM cells. They concluded based on  
13          their results that the attachment and outgrowth of cultured ICM cells is enhanced  
14          by the use of STO (mouse fibroblast) feeder cells (instead of bovine uterus  
15          epithelial cells) and by the use of charcoal-stripped serum (rather than normal se-  
16          rum) to supplement the culture medium. Van Stekelenburg et al reported,  
17          however, that their cell lines resembled epithelial cells more than pluripotent  
18          ICM cells. (*Id.*)

19          Still further, Smith et al., WO 94/24274, published October 27, 1994,  
20          Evans et al, WO 90/03432, published April 5, 1990, and Wheeler et al, WO  
21          94/26889, published November 24, 1994, report the isolation, selection and  
22          propagation of animal stem cells which purportedly may be used to obtain  
23          transgenic animals. Also, Evans et al., WO 90/03432, published on April 5,  
24          1990, reported the derivation of purportedly pluripotent embryonic stem cells  
25          derived from porcine and bovine species which assertedly are useful for the  
26          production of transgenic animals. Further, Wheeler et al, WO 94/26884,

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1 published November 24, 1994, disclosed embryonic stem cells which are  
2 assertedly useful for the manufacture of chimeric and transgenic ungulates.  
3 Thus, based on the foregoing, it is evident that many groups have attempted to  
4 produce ES cell lines, e.g., because of their potential application in the  
5 production of cloned or transgenic embryos and in nuclear transplantation.

6 The use of ungulate ICM cells for nuclear transplantation has also been  
7 reported. For example, Collas et al., *Mol. Reprod. Dev.*, 38:264-267 (1994)  
8 disclose nuclear transplantation of bovine ICMs by microinjection of the lysed  
9 donor cells into enucleated mature oocytes. The reference disclosed culturing of  
10 embryos *in vitro* for seven days to produce fifteen blastocysts which, upon  
11 transferral into bovine recipients, resulted in four pregnancies and two births.  
12 Also, Keefer et al., *Biol. Reprod.*, 50:935-939 (1994), disclose the use of bovine  
13 ICM cells as donor nuclei in nuclear transfer procedures, to produce blastocysts  
14 which, upon transplantation into bovine recipients, resulted in several live  
15 offspring. Further, Sims et al., *Proc. Natl. Acad. Sci., USA*, 90:6143-6147  
16 (1993), disclosed the production of calves by transfer of nuclei from short-term  
17 *in vitro* cultured bovine ICM cells into enucleated mature oocytes.

18 Also, the production of live lambs following nuclear transfer of cultured  
19 embryonic disc cells has been reported (Campbell et al., *Nature*, 380:64-68  
20 (1996)). Still further, the use of bovine pluripotent embryonic cells in nuclear  
21 transfer and the production of chimeric fetuses has also been reported (Stice  
22 et al., *Biol. Reprod.*, 54:100-110 (1996)); Collas et al., *Mol. Reprod. Dev.*,  
23 38:264-267 (1994).

24 Also, there have been previous attempts to produce cross species NT units  
25 (Wolfe et al., *Theriogenology*, 33:350 (1990)). Specifically, bovine embryonic  
26 cells were fused with bison oocytes to produce some cross species NT units

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1 possibly having an inner cell mass. However, embryonic cells, not adult cells  
2 were used, as donor nuclei in the nuclear transfer procedure. The dogma has  
3 been that embryonic cells are more easily reprogrammed than adult cells. This  
4 dates back to earlier NT studies in the frog (review by DiBerardino,  
5 *Differentiation*, 17:17-30 (1980)). Also, this study involved very  
6 phylogenetically similar animals (cattle nuclei and bison oocytes). By contrast,  
7 previously when more diverse species were fused during NT (cattle nuclei into  
8 hamster oocytes), no inner cell mass structures were obtained. Further, it has  
9 never been previously reported that the inner cell mass cells from NT units could  
10 be used to form an ES cell-like colony that could be propagated.

11 Also, Collas et al (*Id.*), taught the use of granulosa cells (adult somatic  
12 cells) to produce bovine nuclear transfer embryos. However, unlike the present  
13 invention, these experiments did not involve cross-species nuclear transfer.  
14 Also, unlike the present invention ES-like cell colonies were not obtained.

15 Very recently, U.S. Patent No. 5,843,780, issued to James A. Thomson  
16 on December 1, 1998, assigned to the Wisconsin Alumni Research Foundation,  
17 that purports to disclose a purified preparation of primate embryonic stem cells  
18 that are (i) capable of proliferation in an *in vitro* culture for over one year; (ii)  
19 maintain a karyotype in which all chromosomes characteristic of the primate  
20 species are present and not noticeably altered through prolonged culture; (iii)  
21 maintains the potential to differentiate into derivatives of endoderm, mesoderm  
22 and ectoderm tissues throughout culture; and (iv) will not differentiate when  
23 cultured on a fibroblast feeder layer. These cells were reportedly negative for the  
24 SSEA-1 marker, positive for the SEA-3 marker, positive for the SSEA-4 marker,  
25 express alkaline phosphatase activity, are pluripotent, and have karyotypes which  
26 include the presence of all the chromosomes characteristic of the primate species



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1 and in which none of the chromosomes are altered. Further, these cells are  
2 respectfully positive for the TRA-1-60, and TRA-1-81 markers. The cells  
3 purportedly differentiate into endoderm, mesoderm and ectoderm cells when  
4 injected into a SCID mouse. Also, purported embryonic stem cell lines derived  
5 from human or primate blastocytes are discussed in Thomson et al., Science  
6 282:1145-1147 and Proc. Natl. Acad. Sci., USA 92:7844-7848 (1995).

7 Thus, Thomson disclose what purportedly are non-human primate and  
8 human embryonic or stem-like cells and methods for their production. However,  
9 there still exists a significant need for methods for producing human embryonic  
10 or stem-like cells that are autologous to an intended transplant recipient given  
11 their significant therapeutic and diagnostic potential.

12 In this regard, numerous human diseases have been identified which may  
13 be treated by cell transplantation. For example, Parkinson's disease is caused by  
14 degeneration of dopaminergic neurons in the substantia nigra. Standard  
15 treatment for Parkinson's involves administration of L-DOPA, which temporarily  
16 ameliorates the loss of dopamine, but causes severe side effects and ultimately  
17 does not reverse the progress of the disease. A different approach to treating  
18 Parkinson's, which promises to have broad applicability to treatment of many  
19 brain diseases and central nervous system injury, involves transplantation of cells  
20 or tissues from fetal or neonatal animals into the adult brain. Fetal neurons from  
21 a variety of brain regions can be incorporated into the adult brain. Such grafts  
22 have been shown to alleviate experimentally induced behavioral deficits, includ-  
23 ing complex cognitive functions, in laboratory animals. Initial test results from  
24 human clinical trials have also been promising. However, supplies of human  
25 fetal cells or tissue obtained from miscarriages is very limited. Moreover,  
26 obtaining cells or tissues from aborted fetuses is highly controversial.

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1           There is currently no available procedure for producing "fetal-like" cells  
2           from the patient. Further, allograft tissue is not readily available and both allo-  
3           graft and xenograft tissue are subject to graft rejection. Moreover, in some cases,  
4           it would be beneficial to make genetic modifications in cells or tissues before  
5           transplantation. However, many cells or tissues wherein such modification  
6           would be desirable do not divide well in culture and most types of genetic  
7           transformation require rapidly dividing cells.

8           There is therefore a clear need in the art for a supply of human embryonic  
9           or stem-like undifferentiated cells for use in transplants and cell and gene thera-  
10          pies.

### 11                           OBJECTS OF THE INVENTION

12           It is an object of the invention to provide novel and improved methods for  
13           producing embryonic or stem-like cells.

14           It is a more specific object of the invention to provide a novel method for  
15           producing embryonic or stem-like cells which involves transplantation of the  
16           nucleus of a mammalian or human cell into an enucleated oocyte of a different  
17           species.

18           It is another specific object of the invention to provide a novel method for  
19           producing non-human primate or human embryonic or stem-like cells which  
20           involves transplantation of the nucleus of a non-human primate or human cell  
21           into an enucleated animal or human oocyte, e.g., an ungulate, human or primate  
22           enucleated oocyte.

23           It is another object of the invention to provide a novel method for  
24           producing lineage-defective non-human primate or human embryonic or stem-  
25           like cells which involves transplantation of the nucleus of a non-human primate  
26           or human cell, e.g., a human adult cell into an enucleated non-human primate or

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1 human oocyte, wherein such cell has been genetically engineered to be incapable  
2 of differentiation into a specific cell lineage or has been modified such that the  
3 cells are "mortal", and thereby do not give rise to a viable offspring, e.g., by  
4 engineering expression of anti-sense or ribozyme telomerase gene.

5 It is still another object of the invention to enhance efficiency of nuclear  
6 transfer and specifically to enhance the development of preimplantation embryos  
7 produced by nuclear transfer by genetically engineering donor somatic cells used  
8 for nuclear transfer to provide for the expression of genes that enhance  
9 embryonic development, e.g., genes of the MHC I family, and in particular Ped  
10 genes such as Q7 and/or Q9.

11 It is yet another object of the invention to enhance the production of  
12 nuclear transfer embryos by IVP and more specifically nuclear transfer embryos  
13 by genetically altering the donor cell used for nuclear transfer such that it is  
14 resistant to apoptosis, e.g., by introduction of a DNA construct that provides for  
15 the expression of genes that inhibit apoptosis, e.g., Bcl-2 or Bcl-2 family  
16 members and/or by the expression of antisense ribozymes specific to genes that  
17 induce apoptosis during early embryonic development.

18 It is still another object of the invention to improve the efficacy of nuclear  
19 transfer by improved selection of donor cells of a specific cell cycle stage, e.g.,  
20 G1 phase, by genetically engineering donor cells such that they express a DNA  
21 construct encoding a particular cyclin linked to a detectable marker, e.g., one that  
22 encodes a visualizable (e.g., fluorescent tag) marker protein.

23 It is also an object of the invention to enhance the development of *in vitro*  
24 produced embryos, by culturing such embryos in the presence of one or more  
25 protease inhibitors, preferably one or more caspase inhibitors, thereby inhibiting  
26 apoptosis.

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1           It is another object of the invention to provide embryonic or stem-like  
2 cells produced by transplantation of nucleus of an animal or human cell into an  
3 enucleated oocyte of a different species.

4           It is a more specific object of the invention to provide primate or human  
5 embryonic or stem-like cells produced by transplantation of the nucleus of a  
6 primate or human cell into an enucleated animal oocyte, e.g., a human, primate  
7 or ungulate enucleated oocyte.

8           It is another object of the invention to use such embryonic or stem-like  
9 cells for therapy or diagnosis.

10          It is a specific object of the invention to use such primate or human  
11 embryonic or stem-like cells for treatment or diagnosis of any disease wherein  
12 cell, tissue or organ transplantation is therapeutically or diagnostically beneficial.

13          It is another specific object of the invention to use the embryonic or stem-  
14 like cells produced according to the invention for the production of differentiated  
15 cells, tissues or organs.

16          It is a more specific object of the invention to use the primate or human  
17 embryonic or stem-like cells produced according to the invention for the  
18 production of differentiated human cells, tissues or organs.

19          It is another specific object of the invention to use the embryonic or stem-  
20 like cells produced according to the invention for the production of genetically  
21 engineered embryonic or stem-like cells, which cells may be used to produce  
22 genetically engineered or transgenic differentiated human cells, tissues or organs,  
23 e.g., having use in gene therapies.

24          It is another specific object of the invention to use the embryonic or stem-  
25 like cells produced according to the invention *in vitro*, e.g. for study of cell dif-  
26 ferentiation and for assay purposes, e.g. for drug studies.

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1           It is another object of the invention to provide improved methods of  
2           transplantation therapy, comprising the usage of isogenic or synegenic cells,  
3           tissues or organs produced from the embryonic or stem-like cells produced  
4           according to the invention. Such therapies include by way of example treatment  
5           of diseases and injuries including Parkinson's, Huntington's, Alzheimer's, ALS,  
6           spinal cord injuries, multiple sclerosis, muscular dystrophy, diabetes, liver  
7           diseases, heart disease, cartilage replacement, burns, vascular diseases, urinary  
8           tract diseases, as well as for the treatment of immune defects, bone marrow trans-  
9           plantation, cancer, among other diseases.

10           It is another object of the invention to use the transgenic or genetically  
11           engineered embryonic or stem-like cells produced according to the invention for  
12           gene therapy, in particular for the treatment and/or prevention of the diseases and  
13           injuries identified, *supra*.

14           It is another object of the invention to use the embryonic or stem-like cells  
15           produced according to the invention or transgenic or genetically engineered  
16           embryonic or stem-like cells produced according to the invention as nuclear  
17           donors for nuclear transplantation.

18           It is still another object of the invention to use genetically engineered ES  
19           cells produced according to the invention for the production of transgenic  
20           animals, e.g., non-human primates, rodents, ungulates, etc. Such transgenic  
21           animals can be used to produce, e.g., animal models for study of human diseases,  
22           or for the production of desired polypeptides, e.g., therapeutics or  
23           nutripharmaceuticals.

24           With the foregoing and other objects, advantages and features of the  
25           invention that will become hereinafter apparent, the nature of the invention may  
26           be more clearly understood by reference to the following detailed description of

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1 the preferred embodiments of the invention and to the appended claims.

2 **BRIEFS DESCRIPTION OF THE FIGURES**

3 Figure 1 is a photograph of a nuclear transfer (NT) unit produced by  
4 transfer of an adult human cell into an enucleated bovine oocyte.

5 Figures 2 to 5 are photographs of embryonic stem-like cells derived from  
6 a NT unit such as is depicted in Figure 1.

7 **DETAILED DESCRIPTION OF THE INVENTION**

8 The present invention provides a novel method for producing embryonic  
9 or stem-like cells, and more specifically non-human primate or human  
10 embryonic or stem-like cells by nuclear transfer or nuclear transplantation. In  
11 the subject application, nuclear transfer or nuclear transplantation or NT are used  
12 interchangeably.

13 As discussed *supra*, the isolation of actual embryonic or stem-like cells  
14 by nuclear transfer or nuclear transplantation has never been reported. Rather,  
15 previous reported isolation of ES-like cells has been from fertilized embryos.  
16 Also, successful nuclear transfer involving cells or DNA of genetically dissimilar  
17 species, or more specifically adult cells or DNA of one species (e.g., human) and  
18 oocytes of another non-related species has never been reported. Rather, while  
19 embryos produced by fusion of cells of closely related species, has been reported,  
20 e.g., bovine-goat and bovine-bison, they did not produce ES cells. (Wolfe et al,  
21 Theriogenology, 33(1):350 (1990).) Also, there has never been reported a  
22 method for producing primate or human ES cells derived from a non-fetal tissue  
23 source. Rather, the limited human fetal cells and tissues which are currently  
24 available must be obtained or derived from spontaneous abortion tissues and  
25 from aborted fetuses.

26 Also, prior to the present invention, no one obtained embryonic or stem-

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1       like cells by cross-species nuclear transplantation.

2               Quite unexpectedly, the present inventors discovered that human  
3       embryonic or stem-like cells and cell colonies may be obtained by  
4       transplantation of the nucleus of a human cell, e.g., an adult differentiated human  
5       cell, into an enucleated animal oocyte, which is used to produce nuclear transfer  
6       (NT) units, the cells of which upon culturing give rise to human embryonic or  
7       stem-like cells and cell colonies. This result is highly surprising because it is the  
8       first demonstration of effective cross-species nuclear transplantation involving  
9       the introduction of a differentiated donor cell or nucleus into an enucleated  
10      oocyte of a genetically dissimilar species, e.g., the transplantation of cell nuclei  
11      from a differentiated animal or human cell, e.g., adult cell, into the enucleated  
12      egg of a different animal species, to produce nuclear transfer units containing  
13      cells which when cultured under appropriate conditions give rise to embryonic  
14      or stem-like cells and cell colonies.

15             Preferably, the NT units used to produce ES-like cells will be cultured to  
16      a size of at least 2 to 400 cells, preferably 4 to 128 cells, and most preferably to  
17      a size of at least about 50 cells.

18             In the present invention, embryonic or stem-like cells refer to cells  
19      produced according to the present invention. The present application refers to  
20      such cells as stem-like cells rather than stem cells because of the manner in  
21      which they are typically produced, i.e., by cross-species nuclear transfer. While  
22      these cells are expected to possess similar differentiation capacity as normal stem  
23      cells they may possess some insignificant differences because of the manner they  
24      are produced. For example, these stem-like cells may possess the mitochondria  
25      of the oocytes used for nuclear transfer, and thus not behave identically to  
26      conventional embryonic stem cells.

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1           The present discovery was made based on the observation that nuclear  
2           transplantation of the nucleus of an adult human cell, specifically a human  
3           epithelial cell obtained from the oral cavity of a human donor, when transferred  
4           into an enucleated bovine oocyte, resulted in the formation of nuclear transfer  
5           units, the cells of which upon culturing gave rise to human stem-like or embry-  
6           onic cells and human embryonic or stem-like cell colonies. This result has  
7           recently been reproduced by transplantation of keratinocytes from an adult  
8           human into an enucleated bovine oocyte with the successful production of a  
9           blastocyst and ES cell line. Based thereon, it is hypothesized by the present  
10          inventors that bovine oocytes and human oocytes, and likely mammals in general  
11          must undergo maturation processes during embryonic development which are  
12          sufficiently similar or conserved so as to permit the bovine oocyte to function as  
13          an effective substitute or surrogate for a human oocyte. Apparently, oocytes in  
14          general comprise factors, likely proteinaceous or nucleic acid in nature, that  
15          induce embryonic development under appropriate conditions, and these functions  
16          that are the same or very similar in different species. These factors may  
17          comprise material RNAs and/or telomerase.

18          Based on the fact that human cell nuclei can be effectively transplanted  
19          into bovine oocytes, it is reasonable to expect that human cells may be  
20          transplanted into oocytes of other non-related species, e.g., other ungulates as  
21          well as other animals. In particular, other ungulate oocytes should be suitable,  
22          e.g. pigs, sheep, horses, goats, etc. Also, oocytes from other sources should be  
23          suitable, e.g. oocytes derived from other primates, amphibians, rodents, rabbits,  
24          guinea pigs, etc. Further, using similar methods, it should be possible to transfer  
25          human cells or cell nuclei into human oocytes and use the resultant blastocytes  
26          to produce human ES cells.



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1           Therefore, in its broadest embodiment, the present invention involves the  
2           transplantation of an animal or human cell nucleus or animal or human cell into  
3           the enucleated oocyte of an animal species different from the donor nuclei, by  
4           injection or fusion, to produce an NT unit containing cells which may be used to  
5           obtain embryonic or stem-like cells and/or cell cultures. For example, the  
6           invention may involve the transplantation of an ungulate cell nucleus or ungulate  
7           cell into an enucleated oocyte of another species, e.g., another ungulate or non-  
8           ungulate, by injection or fusion, which cells and/or nuclei are combined to  
9           produce NT units and which are cultured under conditions suitable to obtain  
10          multicellular NT units, preferably comprising at least about 2 to 400 cells, more  
11          preferably 4 to 128 cells, and most preferably at least about 50 cells. The cells  
12          of such NT units may be used to produce embryonic or stem-like cells or cell  
13          colonies upon culturing.

14          However, the preferred embodiment of the invention comprises the  
15          production of non-human primate or human embryonic or stem-like cells by  
16          transplantation of the nucleus of a donor human cell or a human cell into an  
17          enucleated human, primate, or non-primate animal oocyte, e.g., an ungulate  
18          oocyte, and in a preferred embodiment a bovine enucleated oocyte.

19          In general, the embryonic or stem-like cells will be produced by a nuclear  
20          transfer process comprising the following steps:

21               (i) obtaining desired human or animal cells to be used as a source of  
22               donor nuclei (which may be genetically altered);

23               (ii) obtaining oocytes from a suitable source, e.g. a mammal and most  
24               preferably a primate or an ungulate source, e.g. bovine,

25               (iii) enucleating said oocytes;

26               (iv) transferring the human or animal cell or nucleus into the enucleated

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1 oocyte of an animal species different than the donor cell or nuclei, e.g., by fusion  
2 or injection;

3 (v) culturing the resultant NT product or NT unit to produce multiple cell  
4 structures; and

5 (vi) culturing cells obtained from said embryos to obtain embryonic or  
6 stem-like cells and stem-like cell colonies.

7 Nuclear transfer techniques or nuclear transplantation techniques are  
8 known in the literature and are described in many of the references cited in the  
9 Background of the Invention. See, in particular, Campbell et al, *Theriogenology*,  
10 43:181 (1995); Collas et al, *Mol. Report Dev.*, 38:264-267 (1994); Keefer et al,  
11 *Biol. Reprod.*, 50:935-939 (1994); Sims et al, *Proc. Natl. Acad. Sci., USA*,  
12 90:6143-6147 (1993); WO 94/26884; WO 94/24274, and WO 90/03432, which  
13 are incorporated by reference in their entirety herein. Also, U.S. Patent Nos.  
14 4,944,384 and 5,057,420 describe procedures for bovine nuclear transplantation.  
15 See, also Cibelli et al, *Science*, Vol. 280:1256-1258 (1998).

16 Human or animal cells, preferably mammalian cells, may be obtained and  
17 cultured by well known methods. Human and animal cells useful in the present  
18 invention include, by way of example, epithelial, neural cells, epidermal cells,  
19 keratinocytes, hematopoietic cells, melanocytes, chondrocytes, lymphocytes (B  
20 and T lymphocytes), other immune cells, erythrocytes, macrophages,  
21 melanocytes, monocytes, mononuclear cells, fibroblasts, cardiac muscle cells,  
22 and other muscle cells, etc. Moreover, the human cells used for nuclear transfer  
23 may be obtained from different organs, e.g., skin, lung, pancreas, liver, stomach,  
24 intestine, heart, reproductive organs, bladder, kidney, urethra and other urinary  
25 organs, etc. These are just examples of suitable donor cells. Suitable donor  
26 cells, i.e., cells useful in the subject invention, may be obtained from any cell or

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1 organ of the body. This includes all somatic or germ cells. Preferably, the donor  
2 cells or nucleus would comprise actively dividing, i.e., non-quiescent, cells as  
3 this has been reported to enhance cloning efficacy. Also preferably, such donor  
4 cells will be in the G1 cell cycle.

5 The resultant blastocytes may be used to obtain embryonic stem cell lines  
6 according to the culturing methods reported by Thomson et al., Science  
7 282:1145-1147 (1998) and Thomson et al., Proc. Natl. Acad. Sci., USA 92:7544-  
8 7848 (1995), incorporated by reference in their entirety herein.

9 In the example which follows the cells used as donors for nuclear transfer  
10 were epithelial cells derived from the oral cavity of a human donor and adult  
11 human keratinocytes. However, as discussed, the disclosed method is applicable  
12 to other human cells or nuclei. Moreover, the cell nuclei may be obtained from  
13 both human somatic and germ cells.

14 It is also possible to arrest donor cells at mitosis before nuclear transfer,  
15 using a suitable technique known in the art. Methods for stopping the cell cycle  
16 at various stages have been thoroughly reviewed in U.S. Patent 5,262,409, which  
17 is herein incorporated by reference. In particular, while cycloheximide has been  
18 reported to have an inhibitory effect on mitosis (Bowen and Wilson (1955) J.  
19 Heredity 45: 3-9), it may also be employed for improved activation of mature  
20 bovine follicular oocytes when combined with electric pulse treatment (Yang et  
21 al. (1992) Biol. Reprod. 42 (Suppl. 1): 117).

22 Zygote gene activation is associated with hyperacetylation of Histone H4.  
23 Trichostatin-A has been shown to inhibit histone deacetylase in a reversible  
24 manner (Adenot et al. Differential H4 acetylation of paternal and maternal  
25 chromatin precedes DNA replication and differential transcriptional activity in  
26 pronuclei of 1-cell mouse embryos. Development (Nov. 1997) 124(22): 4615-

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1 4625; Yoshida et al. Trichostatin A and trapoxin: novel chemical probes for the  
2 role of histone acetylation in chromatin structure and function. Bioessays (May,  
3 1995) 17(5): 423-430), as have other compounds.

4 For instance, butyrate is also believed to cause hyper-acetylations of  
5 histones by inhibiting histone deacetylase. Generally, butyrate appears to modify  
6 gene expression and in almost all cases its addition to cells in culture appears to  
7 arrest cell growth. Use of butyrate in this regard is described in U.S. Patent No.  
8 5,681,718, which is herein incorporated by reference. Thus, donor cells may be  
9 exposed to Trichostatin-A or another appropriate deacetylase inhibitor prior to  
10 fusion, or such a compound may be added to the culture media prior to genome  
11 activation.

12 Additionally, demethylation of DNA is thought to be a requirement for  
13 proper access of transcription factors to DNA regulatory sequences. Global  
14 demethylation of DNA from the eight-cell stage to the blastocyst stage in  
15 preimplantation embryos has previously been described (Stein et al., Mol.  
16 Reprod. & Dev. 47(4): 421-429). Also, Jaenisch et al. (1997) have reported that  
17 5-azacytidine can be used to reduce the level of DNA methylation in cells,  
18 potentially leading to increased access of transcription factors to DNA regulatory  
19 sequences. Accordingly, donor cells may be exposed to 5-azacytidine (5-Aza)  
20 previous to fusion, or 5-Aza may be added to the culture medium from the 8 cell  
21 stage to blastocyst. Alternatively, other known methods for effecting DNA  
22 demethylation may be used.

23 The oocytes used for nuclear transfer may be obtained from animals  
24 including mammals and amphibians. Suitable mammalian sources for oocytes  
25 include sheep, bovines, ovines, pigs, horses, rabbits, goats, guinea pigs, mice,  
26 hamsters, rats, primates, humans, etc. In the preferred embodiments, the oocytes

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1 will be obtained from primates or ungulates, e.g., a bovine.

2 Methods for isolation of oocytes are well known in the art. Essentially,  
3 this will comprise isolating oocytes from the ovaries or reproductive tract of a  
4 mammal or amphibian, e.g., a bovine. A readily available source of bovine  
5 oocytes is slaughterhouse materials.

6 For the successful use of techniques such as genetic engineering, nuclear  
7 transfer and cloning, oocytes must generally be matured *in vitro* before these  
8 cells may be used as recipient cells for nuclear transfer, and before they can be  
9 fertilized by the sperm cell to develop into an embryo. This process generally  
10 requires collecting immature (prophase I) oocytes from animal ovaries, e.g.,  
11 bovine ovaries obtained at a slaughterhouse and maturing the oocytes in a  
12 maturation medium prior to fertilization or enucleation until the oocyte attains  
13 the metaphase II stage, which in the case of bovine oocytes generally occurs  
14 about 18-24 hours post-aspiration. For purposes of the present invention, this  
15 period of time is known as the "maturation period." As used herein for  
16 calculation of time periods, "aspiration" refers to aspiration of the immature  
17 oocyte from ovarian follicles.

18 Additionally, metaphase II stage oocytes, which have been matured *in*  
19 *vivo* have been successfully used in nuclear transfer techniques. Essentially,  
20 mature metaphase II oocytes are collected surgically from either non-superovu-  
21 lated or superovulated cows or heifers 35 to 48 hours past the onset of estrus or  
22 past the injection of human chorionic gonadotropin (hCG) or similar hormone.

23 The stage of maturation of the oocyte at enucleation and nuclear transfer  
24 has been reported to be significant to the success of NT methods. (See e.g.,  
25 Prather et al., *Differentiation*, 48, 1-8, 1991). In general, previous successful  
26 mammalian embryo cloning practices use the metaphase II stage oocyte as the

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1 recipient oocyte because at this stage it is believed that the oocyte can be or is  
2 sufficiently "activated" to treat the introduced nucleus as it does a fertilizing  
3 sperm. In domestic animals, and especially cattle, the oocyte activation period  
4 generally ranges from about 16-52 hours, preferably about 28-42 hours post-  
5 aspiration.

6 For example, immature oocytes may be washed in HEPES buffered  
7 hamster embryo culture medium (HECM) as described in Seshagine et al., *Biol.*  
8 *Reprod.*, 40, 544-606, 1989, and then placed into drops of maturation medium  
9 consisting of 50 microliters of tissue culture medium (TCM) 199 containing 10%  
10 fetal calf serum which contains appropriate gonadotropins such as luteinizing  
11 hormone (LH) and follicle stimulating hormone (FSH), and estradiol under a  
12 layer of lightweight paraffin or silicon at 39°C.

13 After a fixed time maturation period, which typically will range from  
14 about 10 to 40 hours, and preferably about 16-18 hours, the oocytes will be  
15 enucleated. Prior to enucleation the oocytes will preferably be removed and  
16 placed in HECM containing 1 milligram per milliliter of hyaluronidase prior to  
17 removal of cumulus cells. This may be effected by repeated pipetting through  
18 very fine bore pipettes or by vortexing briefly. The stripped oocytes are then  
19 screened for polar bodies, and the selected metaphase II oocytes, as determined  
20 by the presence of polar bodies, are then used for nuclear transfer. Enucleation  
21 follows.

22 Enucleation may be effected by known methods, such as described in U.S.  
23 Patent No. 4,994,384 which is incorporated by reference herein. For example,  
24 metaphase II oocytes are either placed in HECM, optionally containing 7.5  
25 micrograms per milliliter cytochalasin B, for immediate enucleation, or may be  
26 placed in a suitable medium, for example CR1aa, plus 10% estrus cow serum,

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1 and then enucleated later, preferably not more than 24 hours later, and more  
2 preferably 16-18 hours later.

3 Enucleation may be accomplished microsurgically using a micropipette  
4 to remove the polar body and the adjacent cytoplasm. The oocytes may then be  
5 screened to identify those of which have been successfully enucleated. This  
6 screening may be effected by staining the oocytes with 1 microgram per milliliter  
7 33342 Hoechst dye in HECM, and then viewing the oocytes under ultraviolet  
8 irradiation for less than 10 seconds. The oocytes that have been successfully  
9 enucleated can then be placed in a suitable culture medium.

10 In the present invention, the recipient oocytes will preferably be  
11 enucleated at a time ranging from about 10 hours to about 40 hours after the  
12 initiation of *in vitro* maturation, more preferably from about 16 hours to about  
13 24 hours after initiation of *in vitro* maturation, and most preferably about 16-18  
14 hours after initiation of *in vitro* maturation.

15 A single animal or human cell or nucleus derived therefrom which is  
16 typically heterologous to the enucleated oocyte will then be transferred into the  
17 perivitelline space of the enucleated oocyte used to produce the NT unit. The  
18 animal or human cell or nucleus and the enucleated oocyte will be used to  
19 produce NT units according to methods known in the art. For example, the cells  
20 may be fused by electrofusion. Electrofusion is accomplished by providing a  
21 pulse of electricity that is sufficient to cause a transient break down of the plasma  
22 membrane. This breakdown of the plasma membrane is very short because the  
23 membrane reforms rapidly. Essentially, if two adjacent membranes are induced  
24 to break down, upon reformation the lipid bilayers intermingle and small chan-  
25 nels will open between the two cells. Due to the thermodynamic instability of  
26 such a small opening, it enlarges until the two cells become one. Reference is

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1 made to U.S. Patent 4,997,384 by Prather et al., (incorporated by reference in its  
2 entirety herein) for a further discussion of this process. A variety of electro-  
3 fusion media can be used including e.g., sucrose, mannitol, sorbitol and phos-  
4 phate buffered solution. Fusion can also be accomplished using Sendai virus as  
5 a fusogenic agent (Graham, *Wister Inot. Symp. Monogr.*, 9, 19, 1969).

6 Also, in some cases (e.g. with small donor nuclei) it may be preferable to  
7 inject the nucleus directly into the oocyte rather than using electroporation  
8 fusion. Such techniques are disclosed in Collas and Barnes, *Mol. Reprod. Dev.*,  
9 38:264-267 (1994), and incorporated by reference in its entirety herein.

10 Preferably, the human or animal cell and oocyte are electrofused in a 500  
11  $\mu$ m chamber by application of an electrical pulse of 90-120V for about 15  $\mu$ sec,  
12 about 24 hours after initiation of oocyte maturation. After fusion, the resultant  
13 fused NT units are then placed in a suitable medium until activation, e.g., one  
14 identified *infra*. Typically activation will be effected shortly thereafter, typically  
15 less than 24 hours later, and preferably about 4-9 hours later.

16 The NT unit may be activated by known methods. Such methods include,  
17 e.g., culturing the NT unit at sub-physiological temperature, in essence by  
18 applying a cold, or actually cool temperature shock to the NT unit. This may be  
19 most conveniently done by culturing the NT unit at room temperature, which is  
20 cold relative to the physiological temperature conditions to which embryos are  
21 normally exposed.

22 Alternatively, activation may be achieved by application of known  
23 activation agents. For example, penetration of oocytes by sperm during  
24 fertilization has been shown to activate prefusion oocytes to yield greater  
25 numbers of viable pregnancies and multiple genetically identical calves after  
26 nuclear transfer. Also, treatments such as electrical and chemical shock or



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1 cycloheximide treatment may also be used to activate NT embryos after fusion.  
2 Suitable oocyte activation methods are the subject of U.S. Patent No. 5,496,720,  
3 to Susko-Parrish et al., which is herein incorporated by reference.

4 For example, oocyte activation may be effected by simultaneously or  
5 sequentially:

- 6 (i) increasing levels of divalent cations in the oocyte, and  
7 (ii) reducing phosphorylation of cellular proteins in the oocyte.

8 This will generally be effected by introducing divalent cations into the  
9 oocyte cytoplasm, e.g., magnesium, strontium, barium or calcium, e.g., in the  
10 form of an ionophore. Other methods of increasing divalent cation levels include  
11 the use of electric shock, treatment with ethanol and treatment with caged  
12 chelators.

13 Phosphorylation may be reduced by known methods, e.g., by the addition  
14 of kinase inhibitors, e.g., serine-threonine kinase inhibitors, such as 6-dimethyl-  
15 amino-purine, staurosporine, 2-aminopurine, and sphingosine.

16 Alternatively, phosphorylation of cellular proteins may be inhibited by  
17 introduction of a phosphatase into the oocyte, e.g., phosphatase 2A and  
18 phosphatase 2B.

19 Activated NT units may be cultured in a suitable *in vitro* culture medium  
20 until the generation of embryonic or stem-like cells and cell colonies. Culture  
21 media suitable for culturing and maturation of embryos are well known in the art.  
22 Examples of known media, which may be used for bovine embryo culture and  
23 maintenance, include Ham's F-10 + 10% fetal calf serum (FCS), Tissue Culture  
24 Medium-199 (TCM-199) + 10% fetal calf serum, Tyrodes-Albumin-Lactate-  
25 Pyruvate (TALP), Dulbecco's Phosphate Buffered Saline (PBS), Eagle's and  
26 Whitten's media. One of the most common media used for the collection and

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1 maturation of oocytes is TCM-199, and 1 to 20% serum supplement including  
2 fetal calf serum, newborn serum, estrual cow serum, lamb serum or steer serum.  
3 A preferred maintenance medium includes TCM-199 with Earl salts, 10% fetal  
4 calf serum, 0.2 MM Ma pyruvate and 50 µg/ml gentamicin sulphate. Any of the  
5 above may also involve co-culture with a variety of cell types such as granulosa  
6 cells, oviduct cells, BRL cells and uterine cells and STO cells.

7 In particular, human epithelial cells of the endometrium secrete leukemia  
8 inhibitory factor (LIF) during the preimplantation and implantation period.  
9 Therefore, the addition of LIF to the culture medium could be of importance in  
10 enhancing the *in vitro* development of the reconstructed embryos. The use of  
11 LIF for embronic or stem-like cell cultures has been described in U.S. Patent  
12 5,712,156, which is herein incorporated by reference.

13 Another maintenance medium is described in U.S. Patent 5,096,822 to  
14 Rosenkrans, Jr. et al., which is incorporated herein by reference. This embryo  
15 medium, named CR1, contains the nutritional substances necessary to support an  
16 embryo. CR1 contains hemicalcium L-lactate in amounts ranging from 1.0 mM  
17 to 10 mM, preferably 1.0 mM to 5.0 mM. Hemicalcium L-lactate is L-lactate  
18 with a hemicalcium salt incorporated thereon.

19 Also, suitable culture medium for maintaining human embryonic cells in  
20 culture as discussed in Thomson et al., Science 282:1145-1147 (1998) and Proc.  
21 Natl. Acad. Sci., USA 92:7844-7848 (1995).

22 Afterward, the cultured NT unit or units are preferably washed and then  
23 placed in a suitable media, e.g., CR1aa medium, Ham's F-10, Tissue Culture  
24 Media-199 (TCM-199). Tyrodes-Albumin-Lactate-Pyrovate (TALP) Dulbecco's  
25 Phosphate Buffered Saline (PBS), Eagle's or Whitten's, preferably containing  
26 about 10% FCS. Such culturing will preferably be effected in well plates which

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1 contain a suitable confluent feeder layer. Suitable feeder layers include, by way  
2 of example, fibroblasts and epithelial cells, e.g., fibroblasts and uterine epithelial  
3 cells derived from ungulates, chicken fibroblasts, murine (e.g., mouse or rat)  
4 fibroblasts, STO and SI-m220 feeder cell lines, and BRL cells.

5 In the preferred embodiment, the feeder cells will comprise mouse  
6 embryonic fibroblasts. Means for preparation of a suitable fibroblast feeder layer  
7 are described in the example which follows and is well within the skill of the  
8 ordinary artisan.

9 The NT units are cultured on the feeder layer until the NT units reach a  
10 size suitable for obtaining cells which may be used to produce embryonic stem-  
11 like cells or cell colonies. Preferably, these NT units will be cultured until they  
12 reach a size of at least about 2 to 400 cells, more preferably about 4 to 128 cells,  
13 and most preferably at least about 50 cells. The culturing will be effected under  
14 suitable conditions, i.e., about 38.5°C and 5% CO<sub>2</sub>, with the culture medium  
15 changed in order to optimize growth typically about every 2-5 days, preferably  
16 about every 3 days.

17 In the case of human cell/enucleated bovine oocyte derived NT units,  
18 sufficient cells to produce an ES cell colony, typically on the order of about 50  
19 cells, will be obtained about 12 days after initiation of oocyte activation.  
20 However, this may vary dependent upon the particular cell used as the nuclear  
21 donor, the species of the particular oocyte, and culturing conditions. One skilled  
22 in the art can readily ascertain visually when a desired sufficient number of cells  
23 has been obtained based on the morphology of the cultured NT units.

24 In the case of human/human nuclear transfer embryos, it may be  
25 advantageous to use culture medium known to be useful for maintaining human  
26 cells in tissue culture. Examples of a culture media suitable for human embryo

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1 culture include the medium reported in Jones et al, *Human Reprod.*, 13(1):169-  
2 177 (1998), the P1-catalog #99242 medium, and the P-1 catalog #99292 medium,  
3 both available from Irvine Scientific, Santa Ana, California, and those used by  
4 Thomson et al. (1998) and (1995). (Id.).

5 As discussed above, the cells used in the present invention will preferably  
6 comprise mammalian somatic cells, most preferably cells derived from an  
7 actively proliferating (non-quiescent) mammalian cell culture. In an especially  
8 preferred embodiment, the donor cell will be genetically modified by the  
9 addition, deletion or substitution of a desired DNA sequence. For example, the  
10 donor cell, e.g., a keratinocyte or fibroblast, e.g., of human, primate or bovine  
11 origin, may be transfected or transformed with a DNA construct that provides for  
12 the expression of a desired gene product, e.g., therapeutic polypeptide. Examples  
13 thereof include lymphokines, e.g., IGF-I, IGF-II, interferons, colony stimulating  
14 factors, connective tissue polypeptides such as collagens, genetic factors, clotting  
15 factors, enzymes, enzyme inhibitors, etc.

16 Also, as discussed above, the donor cells may be modified prior to nuclear  
17 transfer to achieve other desired effects, e.g., impaired cell lineage development,  
18 enhanced embryonic development and/or inhibition of apoptosis. Examples of  
19 desirable modifications are discussed further below.

20 One aspect of the invention will involve genetic modification of the donor  
21 cell, e.g., a human cell, such that it is lineage deficient and therefore when used  
22 for nuclear transfer it will be unable to give rise to a viable offspring. This is  
23 desirable especially in the context of human nuclear transfer embryos, wherein  
24 for ethical reasons, production of a viable embryo may be an unwanted outcome.  
25 This can be effected by genetically engineering a human cell such that it is  
26 incapable of differentiating into specific cell lineages when used for nuclear

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1 transfer. In particular, cells may be genetically modified such that when used as  
2 nuclear transfer donors the resultant "embryos" do not contain or substantially  
3 lack at least one of mesoderm, endoderm or ectoderm tissue.

4 It is anticipated that this can be accomplished by knocking out or  
5 impairing the expression of one or more mesoderm, endoderm or ectoderm  
6 specific genes. Examples thereof include:

7 Mesoderm: SRF, MESP-1, HNF-4, beta-I integrin, MSD;

8 Endoderm: GATA-6, GATA-4;

9 Ectoderm: RNA helicase A, H beta 58.

10 The above list is intended to be exemplary and non-exhaustive of known  
11 genes which are involved in the development of mesoderm, endoderm and  
12 ectoderm. The generation of mesoderm deficient, endoderm deficient and  
13 ectoderm deficient cells and embryos has been previously reported in the  
14 literature. See, e.g., Arsenian et al, *EMBO J.*, Vol. 17(2):6289-6299 (1998); Saga  
15 Y, *Mech. Dev.*, Vol. 75(1-2):53-66 (1998); Holdener et al, *Development*, Voll  
16 120(5):1355-1346 (1994); Chen et al, *Genes Dev.* Vol. 8(20):2466-2477 (1994);  
17 Rohwedel et al, *Dev. Biol.*, 201(2):167-189 (1998) (mesoderm); Morrissey et al,  
18 *Genes, Dev.*, Vol. 12(22):3579-3590 (1998); Soudais et al, *Development*, Vol.  
19 121(11):3877-3888 (1995) (endoderm); and Lee et al, *Proc. Natl. Acad. Sci.*  
20 *USA*, Vol. 95:(23):13709-13713 (1998); and Radice et al, *Development*, Vol.  
21 111(3):801-811 (1991) (ectoderm).

22 In general, a desired somatic cell, e.g., a human keratinocyte, epithelial  
23 cell or fibroblast, will be genetically engineered such that one or more genes  
24 specific to particular cell lineages are "knocked out" and/or the expression of  
25 such genes significantly impaired. This may be effected by known methods, e.g.,  
26 homologous recombination. A preferred genetic system for effecting "knock-

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1 out" of desired genes is disclosed by Capecchi et al, U.S. Patents 5,631,153 and  
2 5,464,764, which reports positive-negative selection (PNS) vectors that enable  
3 targeted modification of DNA sequences in a desired mammalian genome. Such  
4 genetic modification will result in a cell that is incapable of differentiating into  
5 a particular cell lineage when used as a nuclear transfer donor.

6 This genetically modified cell will be used to produce a lineage-defective  
7 nuclear transfer embryo, i.e., that does not develop at least one of a functional  
8 mesoderm, endoderm or ectoderm. Thereby, the resultant embryos, even if  
9 implanted, e.g., into a human uterus, would not give rise to a viable offspring.  
10 However, the ES cells that result from such nuclear transfer will still be useful  
11 in that they will produce cells of the one or two remaining non-impaired lineage.  
12 For example, an ectoderm deficient human nuclear transfer embryo will still give  
13 rise to mesoderm and endoderm derived differentiated cells. An ectoderm  
14 deficient cell can be produced by deletion and/or impairment of one or both of  
15 RNA helicase A or H beta 58 genes.

16 These lineage deficient donor cells may also be genetically modified to  
17 express another desired DNA sequence.

18 Thus, the genetically modified donor cell will give rise to a lineage-  
19 deficient blastocyst which, when plated, will differentiate into at most two of the  
20 embryonic germ layers.

21 Alternatively, the donor cell can be modified such that it is "mortal". This  
22 can be achieved by expressing anti-sense or ribozyme telomerase genes. This  
23 can be effected by known genetic methods that will provide for expression of  
24 antisense DNA or ribozymes, or by gene knockout. These "mortal" cells, when  
25 used for nuclear transfer, will not be capable of differentiating into viable  
26 offspring.

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1           Another preferred embodiment of the present invention is the production  
2 of nuclear transfer embryos that grow more efficiently in tissue culture. This is  
3 advantageous in that it should reduce the requisite time and necessary fusions to  
4 produce ES cells and/or offspring (if the blastocysts are to be implanted into a  
5 female surrogate). This is desirable also because it has been observed that  
6 blastocysts and ES cells resulting from nuclear transfer may have impaired  
7 development potential. While these problems may often be alleviated by  
8 alteration of tissue culture conditions, an alternative solution is to enhance  
9 embryonic development by enhancing expression of genes involved in  
10 embryonic development.

11           For example, it has been reported that the gene products of the Ped type,  
12 which are members of the MHC I family, are of significant importance to  
13 embryonic development. More specifically, it has been reported in the case of  
14 mouse preimplantation embryos that the Q7 and Q9 genes are responsible for the  
15 "fast growth" phenotype. Therefore, it is anticipated that introduction of DNAs  
16 that provide for the expression of these and related genes, or their human or other  
17 mammalian counterparts into donor cells, will give rise to nuclear transfer  
18 embryos that grow more quickly. This is particularly desirable in the context of  
19 cross-species nuclear transfer embryos which may develop less efficiently in  
20 tissue culture than nuclear transfer embryos produced by fusion of cells or nuclei  
21 of the same species.

22           In particular, a DNA construct containing the Q7 and/or Q9 gene will be  
23 introduced into donor somatic cells prior to nuclear transfer. For example, an  
24 expression construct can be constructed containing a strong constitutive  
25 mammalian promoter operably linked to the Q7 and/or Q9 genes, an IRES, one  
26 or more suitable selectable markers, e.g., neomycin, ADA, DHFR, and a poly-A

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1 sequence, e.g., bGH polyA sequence. Also, it may be advantageous to further  
2 enhance Q7 and Q9 gene expression by the inclusion of insulates. It is  
3 anticipated that these genes will be expressed early on in blastocyst development  
4 as these genes are highly conserved in different species, e.g., bovines, goats,  
5 porcine, dogs, cats, and humans. Also, it is anticipated that donor cells can be  
6 engineered to affect other genes that enhance embryonic development. Thus,  
7 these genetically modified donor cells should produce blastocysts and  
8 preimplantation stage embryos more efficiently.

9 Still another aspect of the invention involves the construction of donor  
10 cells that are resistant to apoptosis, i.e., programmed cell death. It has been  
11 reported in the literature that cell death related genes are present in  
12 preimplantation stage embryos. (Adams et al, *Science*, 281(5381):1322-1326  
13 (1998)). Genes reported to induce apoptosis include, e.g., Bad, Bok, BH3, Bik,  
14 Hrk, BNIP3, Bim<sub>L</sub>, Bad, Bid, and EGL-1. By contrast, genes that reportedly  
15 protect cells from programmed cell death include, by way of example, Bcl-XL,  
16 Bcl-w, Mcl-1, A1, Nr-13, BHRF-1, LMW5-HL, ORF16, Ks-Bcl-2, E1B-19K,  
17 and CED-9.

18 Thus, donor cells can be constructed wherein genes that induce apoptosis  
19 are "knocked out" or wherein the expression of genes that protect the cells from  
20 apoptosis is enhanced or turned on during embryonic development.

21 For example, this can be effected by introducing a DNA construct that  
22 provides for regulated expression of such protective genes, e.g., Bcl-2 or related  
23 genes during embryonic development. Thereby, the gene can be "turned on" by  
24 culturing the embryo under specific growth conditions. Alternatively, it can be  
25 linked to a constitutive promoter.

26 More specifically, a DNA construct containing a Bcl-2 gene operably



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1 linked to a regulatable or constitutive promoter, e.g., PGK, SV40, CMV,  
2 ubiquitin, or beta-actin, an IRES, a suitable selectable marker, and a poly-A  
3 sequence can be constructed and introduced into a desired donor mammalian  
4 cell, e.g., human keratinocyte or fibroblast.

5 These donor cells, when used to produce nuclear transfer embryos, should  
6 be resistant to apoptosis and thereby differentiate more efficiently in tissue  
7 culture. Thereby, the speed and/or number of suitable preimplantation embryos  
8 produced by nuclear transfer can be increased.

9 Another means of accomplishing the same result is to impair the  
10 expression of one or more genes that induce apoptosis. This will be effected by  
11 knock-out or by the use of antisense or ribozymes against genes that are  
12 expressed in and which induce apoptosis early on in embryonic development.  
13 Examples thereof are identified above. Still alternatively, donor cells may be  
14 constructed containing both modifications, i.e., impairment of apoptosis-inducing  
15 genes and enhanced expression of genes that impede or prevent apoptosis. The  
16 construction and selection of genes that affect apoptosis, and cell lines that  
17 express such genes, is disclosed in U.S. Patent No. 5,646,008, Craig B.  
18 Thompson et al inventors, and assigned to the University of Michigan. This  
19 patent is incorporated by reference herein.

20 One means of enhancing efficiency is to select cells of a particular cell  
21 cycle stage as the donor cell. It has been reported that this can have significant  
22 effects on nuclear transfer efficiency. (Barnes et al, *Mol. Reprod. Devel.*,  
23 36(1):33-41 (1993). Different methods for selecting cells of a particular cell  
24 cycle stage have been reported and include serum starvation (Campbell et al,  
25 *Nature*, 380:64-66 (1996); Wilmut et al, *Nature*, 385:810-813 (1997), and  
26 chemical synchronization (Urbani et al, *Exp. Cell Res.*, 219(1):159-168 (1995).

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1 For example, a particular cyclin DNA may be operably linked to a regulatory  
2 sequence, together with a detectable marker, e.g., green fluorescent protein  
3 (GFP), followed by the cyclin destruction box, and optionally insulation  
4 sequences to enhance cyclin and marker protein expression. Thereby, cells of a  
5 desired cell cycle can be easily visually detected and selected for use as a nuclear  
6 transfer donor. An example thereof is the cyclin D1 gene in order to select for  
7 cells that are in G1. However, any cyclin gene should be suitable for use in the  
8 claimed invention. (See, e.g., King et al, *Mol. Biol. Cell*, Vol. 7(9):1343-1357  
9 (1996)).

10 However, a less invasive or more efficient method for producing cells of  
11 a desired cell cycle stage are needed. It is anticipated that this can be effected by  
12 genetically modifying donor cells such that they express specific cyclins under  
13 detectable conditions. Thereby, cells of a specific cell cycle can be readily  
14 discerned from other cell cycles.

15 Cyclins are proteins that are expressed only during specific stages of the  
16 cell cycle. They include cyclin D1, D2 and D3 in G1 phase, cyclin B1 and B2  
17 in G2/M phase and cyclin E, A and H in S phase. These proteins are easily  
18 translated and destroyed in the cytosol. This "transient" expression of  
19 such proteins is attributable in part to the presence of a "destruction box", which  
20 is a short amino acid sequence that is part of the protein that functions as a tag  
21 to direct the prompt destruction of these proteins via the ubiquitin pathway.  
22 (Adams et al, *Science*, 281 (5321):1322-1326 (1998)).

23 In the present invention, donor cells will be constructed that express one  
24 or more of such cyclin genes under easily detectable conditions, preferably  
25 visualizable, e.g., by the use of a fluorescent label. For example, a particular  
26 cyclin DNA may be operably linked to a regulatory sequence, together with a

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1 detectable marker, e.g., green fluorescent protein (GFP), followed by the cyclin  
2 destruction box, and optionally insulation sequences to enhance cyclin and/or  
3 marker protein expression. Thereby, cells of a desired cell cycle can be easily  
4 visually detected and selected for use as a nuclear transfer donor. An example  
5 thereof is the cyclin D1 gene which can be used to select for cells that are in G1.  
6 However, any cyclin gene should be suitable for use in the claimed invention.  
7 (*See, e.g., King et al, Mol. Biol. Cell, Vol. 7(9):1343-1357 (1996)*).

8 Still another aspect of the invention is a method for enhancing nuclear  
9 transfer efficiency, preferably in a cross-species nuclear transfer process. While  
10 the present inventors have demonstrated that nuclei or cells of one species when  
11 inserted or fused with an enucleated oocyte of a different species can give rise  
12 to nuclear transfer embryos that produce blastocysts, which embryos can give  
13 rise to ES cell lines, the efficiency of such process is quite low. Therefore, many  
14 fusions typically need to be effected to produce a blastocyst the cells of which  
15 may be cultured to produce ES cells and ES cell lines.

16 Yet another means for enhancing the development of nuclear transfer  
17 embryos *in vitro* is by optimizing culture conditions. One means of achieving  
18 this result will be to culture NT embryos under conditions impede apoptosis.  
19 With respect to this embodiment of the invention, it has been found that  
20 proteases such as capsases can cause oocyte death by apoptosis similar to other  
21 cell types. (*See, Jurisicosva et al, Mol. Reprod. Devel., 51(3):243-253 (1998)*.)

22 It is anticipated that blastocyst development will be enhanced by including  
23 in culture media used for nuclear transfer and to maintain blastocysts or culture  
24 preimplantation stage embryos one or more capsase inhibitors. Such inhibitors  
25 include by way of example capsase-4 inhibitor I, capsase-3 inhibitor I, capsase-6  
26 inhibitor II, capsase-9 inhibitor II, and capsase-1 inhibitor I. the amount thereof

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1 will be an amount effective to inhibit apoptosis, e.g., 0.00001 to 5.0% by weight  
2 of medium; more preferably 0.01% to 1.0% by weight of medium. Thus, the  
3 foregoing methods may be used to increase the efficiency of nuclear transfer by  
4 enhancing subsequent blastocyst and embryo development in tissue culture.

5 After NT units of the desired size are obtained, the cells are mechanically  
6 removed from the zone and are then used to produce embryonic or stem-like  
7 cells and cell lines. This is preferably effected by taking the clump of cells  
8 which comprise the NT unit, which typically will contain at least about 50 cells,  
9 washing such cells, and plating the cells onto a feeder layer, e.g., irradiated fibro-  
10 blast cells. Typically, the cells used to obtain the stem-like cells or cell colonies  
11 will be obtained from the inner most portion of the cultured NT unit which is  
12 preferably at least 50 cells in size. However, NT units of smaller or greater cell  
13 numbers as well as cells from other portions of the NT unit may also be used to  
14 obtain ES-like cells and cell colonies.

15 It may be that a longer exposure of donor cell DNA to the oocyte's  
16 cytosol would facilitate the dedifferentiation process. In this regard, recloning  
17 could be accomplished by taking blastomeres from a reconstructed embryo and  
18 fusing them with a new enucleated oocyte. Alternatively, the donor cell may be  
19 fused with an enucleated oocyte and 4 to 6 hours later, without activation,  
20 chromosomes may be removed and fused with a younger oocyte. Activation  
21 would occur thereafter.

22 The cells are maintained in the feeder layer in a suitable growth medium,  
23 e.g., alpha MEM supplemented with 10% FCS and 0.1 mM beta-  
24 mercaptoethanol (Sigma) and L-glutamine. The growth medium is changed as  
25 often as necessary to optimize growth, e.g., about every 2-3 days.

26 This culturing process results in the formation of embryonic or stem-like

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1 cells or cell lines. In the case of human cell/bovine oocyte derived NT embryos,  
2 colonies are observed by about the second day of culturing in the alpha MEM  
3 medium. However, this time may vary dependent upon the particular nuclear  
4 donor cell, specific oocyte and culturing conditions. One skilled in the art can  
5 vary the culturing conditions as desired to optimize growth of the particular  
6 embryonic or stem-like cells.

7 The embryonic or stem-like cells and cell colonies obtained will typically  
8 exhibit an appearance similar to embryonic or stem-like cells of the species used  
9 as the nuclear cell donor rather than the species of the donor oocyte. For  
10 example, in the case of embryonic or stem-like cells obtained by the transfer of  
11 a human nuclear donor cell into an enucleated bovine oocyte, the cells exhibit a  
12 morphology more similar to mouse embryonic stem cells than bovine ES-like  
13 cells.

14 More specifically, the individual cells of the human ES-line cell colony  
15 are not well defined, and the perimeter of the colony is refractive and smooth in  
16 appearance. Further, the cell colony has a longer cell doubling time, about twice  
17 that of mouse ES cells. Also, unlike bovine and porcine derived ES cells, the  
18 colony does not possess an epithelial-like appearance.

19 As discussed above, it has been reported by Thomson, in U.S. Patent  
20 5,843,780, that primate stem cells are SSEA-1 (-), SSEA-4 (+), TRA-1-60 (+),  
21 TRA-1-81 (+) and alkaline phosphatase (+). It is anticipated that human and  
22 primate ES cells produced according to the present methods will exhibit similar  
23 or identical marker expression.

24 Alternatively, that such cells are actual human or primate embryonic stem  
25 cells will be confirmed based on their capability of giving rise to all of  
26 mesoderm, ectoderm and endoderm tissues. This will be demonstrated by

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1 culturing ES cells produced according to the invention under appropriate  
2 conditions, e.g., as disclosed by Thomsen, U.S. Patent 5,843,780, incorporated  
3 by reference in its entirety herein.

4 The resultant embryonic or stem-like cells and cell lines, preferably  
5 human embryonic or stem-like cells and cell lines, have numerous therapeutic  
6 and diagnostic applications. Most especially, such embryonic or stem-like cells  
7 may be used for cell transplantation therapies. Human embryonic or stem-like  
8 cells have application in the treatment of numerous disease conditions.

9 In this regard, it is known that mouse embryonic stem (ES) cells are  
10 capable of differentiating into almost any cell type, e.g., hematopoietic stem  
11 cells. Therefore, human embryonic or stem-like cells produced according to the  
12 invention should possess similar differentiation capacity. The embryonic or  
13 stem-like cells according to the invention will be induced to differentiate to  
14 obtain the desired cell types according to known methods. For example, the  
15 subject human embryonic or stem-like cells may be induced to differentiate into  
16 hematopoietic stem cells, muscle cells, cardiac muscle cells, liver cells, cartilage  
17 cells, epithelial cells, urinary tract cells, etc., by culturing such cells in  
18 differentiation medium and under conditions which provide for cell differentia-  
19 tion. Medium and methods which result in the differentiation of embryonic stem  
20 cells are known in the art as are suitable culturing conditions.

21 For example, Palacios et al, *Proc. Natl. Acad. Sci., USA*, 92:7530-7537  
22 (1995) teaches the production of hematopoietic stem cells from an embryonic  
23 cell line by subjecting stem cells to an induction procedure comprising initially  
24 culturing aggregates of such cells in a suspension culture medium lacking  
25 retinoic acid followed by culturing in the same medium containing retinoic acid,  
26 followed by transferral of cell aggregates to a substrate which provides for cell

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1 attachment.

2 Moreover, Pedersen, *J. Reprod. Fertil. Dev.*, 6:543-552 (1994) is a review  
3 article which references numerous articles disclosing methods for *in vitro*  
4 differentiation of embryonic stem cells to produce various differentiated cell  
5 types including hematopoietic cells, muscle, cardiac muscle, nerve cells, among  
6 others.

7 Further, Bain et al, *Dev. Biol.*, 168:342-357 (1995) teaches *in vitro*  
8 differentiation of embryonic stem cells to produce neural cells which possess  
9 neuronal properties. These references are exemplary of reported methods for  
10 obtaining differentiated cells from embryonic or stem-like cells. These  
11 references and in particular the disclosures therein relating to methods for  
12 differentiating embryonic stem cells are incorporated by reference in their  
13 entirety herein.

14 Thus, using known methods and culture medium, one skilled in the art  
15 may culture the subject embryonic or stem-like cells to obtain desired  
16 differentiated cell types, e.g., neural cells, muscle cells, hematopoietic cells, etc.

17 In addition, the use of inducible Bcl-2 or Bcl-xl might be useful for  
18 enhancing *in vitro* development of specific cell lineages. *In vivo*, Bcl-2 prevents  
19 many, but not all, forms of apoptotic cell death that occur during lymphoid and  
20 neural development. A thorough discussion of how Bcl-2 expression might be  
21 used to inhibit apoptosis of relevant cell lineages following transfection of donor  
22 cells is disclosed in U.S. Patent No. 5,646,008, which is herein incorporated by  
23 reference.

24 The subject embryonic or stem-like cells may be used to obtain any  
25 desired differentiated cell type. Therapeutic usages of such differentiated human  
26 cells are unparalleled. For example, human hematopoietic stem cells may be

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1        used in medical treatments requiring bone marrow transplantation. Such  
2        procedures are used to treat many diseases, e.g., late stage cancers such as  
3        ovarian cancer and leukemia, as well as diseases that compromise the immune  
4        system, such as AIDS. Hematopoietic stem cells can be obtained, e.g., by fusing  
5        adult somatic cells of a cancer or AIDS patient, e.g., epithelial cells or lympho-  
6        cytes with an enucleated oocyte, e.g., bovine oocyte, obtaining embryonic or  
7        stem-like cells as described above, and culturing such cells under conditions  
8        which favor differentiation, until hematopoietic stem cells are obtained. Such  
9        hematopoietic cells may be used in the treatment of diseases including cancer  
10       and AIDS.

11       Alternatively, adult somatic cells from a patient with a neurological  
12       disorder may be fused with an enucleated animal oocyte, e.g., a primate or  
13       bovine oocyte, human embryonic or stem-like cells obtained therefrom, and such  
14       cells cultured under differentiation conditions to produce neural cell lines.  
15       Specific diseases treatable by transplantation of such human neural cells include,  
16       by way of example, Parkinson's disease, Alzheimer's disease, ALS and cerebral  
17       palsy, among others. In the specific case of Parkinson's disease, it has been  
18       demonstrated that transplanted fetal brain neural cells make the proper connec-  
19       tions with surrounding cells and produce dopamine. This can result in long-term  
20       reversal of Parkinson's disease symptoms.

21       To allow for specific selection of differentiated cells, donor cells may be  
22       transfected with selectable markers expressed via inducible promoters, thereby  
23       permitting selection or enrichment of particular cell lineages when  
24       differentiation is induced. For example, CD34-neo may be used for selection of  
25       hematopoietic cells, Pw1-neo for muscle cells, Mash-1-neo for sympathetic  
26       neurons, Mal-neo for human CNS neurons of the grey matter of the cerebral



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1 cortex, etc.

2 The great advantage of the subject invention is that it provides an  
3 essentially limitless supply of isogenic or syngenic human cells suitable for  
4 transplantation. Therefore, it will obviate the significant problem associated with  
5 current transplantation methods, i.e., rejection of the transplanted tissue which  
6 may occur because of host-vs-graft or graft-vs-host rejection. Conventionally,  
7 rejection is prevented or reduced by the administration of anti-rejection drugs  
8 such as cyclosporine. However, such drugs have significant adverse side-effects,  
9 e.g., immunosuppression, carcinogenic properties, as well as being very  
10 expensive. The present invention should eliminate, or at least greatly reduce, the  
11 need for anti-rejection drugs.

12 Other diseases and conditions treatable by isogenic cell therapy include,  
13 by way of example, spinal cord injuries, multiple sclerosis, muscular dystrophy,  
14 diabetes, liver diseases, i.e., hypercholesterolemia, heart diseases, cartilage  
15 replacement, burns, foot ulcers, gastrointestinal diseases, vascular diseases,  
16 kidney disease, urinary tract disease, and aging related diseases and conditions.

17 Also, human embryonic or stem-like cells produced according to the  
18 invention may be used to produce genetically engineered or transgenic human  
19 differentiated cells. Essentially, this will be effected by introducing a desired  
20 gene or genes, which may be heterologous, or removing all or part of an  
21 endogenous gene or genes of human embryonic or stem-like cells produced  
22 according to the invention, and allowing such cells to differentiate into the de-  
23 sired cell type. A preferred method for achieving such modification is by  
24 homologous recombination because such technique can be used to insert, delete  
25 or modify a gene or genes at a specific cite or cites in the stem-like cell genome.

26 This methodology can be used to replace defective genes, e.g., defective

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1 immune system genes, cystic fibrosis genes, or to introduce genes which result  
2 in the expression of therapeutically beneficial proteins such as growth factors,  
3 lymphokines, cytokines, enzymes, etc. For example, the gene encoding brain  
4 derived growth factor may be introduced into human embryonic or stem-like  
5 cells, the cells differentiated into neural cells and the cells transplanted into a  
6 Parkinson's patient to retard the loss of neural cells during such disease.

7 Previously, cell types transfected with BDNF varied from primary cells  
8 to immortalized cell lines, either neural or non-neural (myoblast and fibroblast)  
9 derived cells. For example, astrocytes have been transfected with BDNF gene  
10 using retroviral vectors, and the cells grafted into a rat model of Parkinson's  
11 disease (Yoshimoto et al., *Brain Research*, 691:25-36, (1995)).

12 This *ex vivo* therapy reduced Parkinson's-like symptoms in the rats up to  
13 45% 32 days after transfer. Also, the tyrosine hydroxylase gene has been placed  
14 into astrocytes with similar results (Lundberg et al., *Develop. Neurol.*, 139:39-53  
15 (1996) and references cited therein).

16 However, such *ex vivo* systems have problems. In particular, retroviral  
17 vectors currently used are down-regulated *in vivo* and the transgene is only  
18 transiently expressed (review by Mulligan, *Science*, 260:926-932 (1993)). Also,  
19 such studies used primary cells, astrocytes, which have finite life span and  
20 replicate slowly. Such properties adversely affect the rate of transfection and  
21 impede selection of stably transfected cells. Moreover, it is almost impossible  
22 to propagate a large population of gene targeted primary cells to be used in  
23 homologous recombination techniques.

24 By contrast, the difficulties associated with retroviral systems should be  
25 eliminated by the use of human embryonic or stem-like cells. It has been demon-  
26 strated previously by the subject assignee that cattle and pig embryonic cell lines

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1 can be transfected and selected for stable integration of heterologous DNA. Such  
2 methods are described in commonly assigned U.S. Serial No. 08/626,054, filed  
3 April 1, 1996, incorporated by reference in its entirety. Therefore, using such  
4 methods or other known methods, desired genes may be introduced into the sub-  
5 ject human embryonic or stem-like cells, and the cells differentiated into desired  
6 cell types, e.g., hematopoietic cells, neural cells, pancreatic cells, cartilage cells,  
7 etc.

8 Genes which may be introduced into the subject embryonic or stem-like  
9 cells include, by way of example, epidermal growth factor, basic fibroblast  
10 growth factor, glial derived neurotrophic growth factor, insulin-like growth  
11 factor (I and II), neurotrophin-3, neurotrophin-4/5, ciliary neurotrophic factor,  
12 AFT-1, cytokine genes (interleukins, interferons, colony stimulating factors,  
13 tumor necrosis factors (alpha and beta), etc.), genes encoding therapeutic  
14 enzymes, collagen, human serum albumin, etc.

15 In addition, it is also possible to use one of the negative selection systems  
16 now known in the art for eliminating therapeutic cells from a patient if necessary.  
17 For example, donor cells transfected with the thymidine kinase (TK) gene will  
18 lead to the production of embryonic cells containing the TK gene.  
19 Differentiation of these cells will lead to the isolation of therapeutic cells of  
20 interest which also express the TK gene. Such cells may be selectively  
21 eliminated at any time from a patient upon gancyclovir administration. Such a  
22 negative selection system is described in U.S. Patent No. 5,698,446, and is herein  
23 incorporated by reference.

24 The subject embryonic or stem-like cells, preferably human cells, also  
25 may be used as an *in vitro* model of differentiation, in particular for the study of  
26 genes which are involved in the regulation of early development.

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Also, differentiated cell tissues and organs using the subject embryonic or stem-like cells may be used in drug studies.

Further, the subject embryonic or stem-like cells may be used as nuclear donors for the production of other embryonic or stem-like cells and cell colonies.

In order to more clearly describe the subject invention, the following examples are provided.

### EXAMPLE 1

#### MATERIALS AND METHODS

##### Donor Cells for Nuclear Transfer

Epithelial cells were lightly scraped from the inside of the mouth of a consenting adult with a standard glass slide. The cells were washed off the slide into a petri dish containing phosphate buffered saline without Ca or Mg. The cells were pipetted through a small-bore pipette to break up cell clumps into a single cell suspension. The cells were then transferred into a microdrop of TL-HEPES medium containing 10% fetal calf serum (FCS) under oil for nuclear transfer into enucleated cattle oocytes.

##### Nuclear Transfer Procedures

Basic nuclear transfer procedures have been described previously. Briefly, after slaughterhouse oocytes were matured *in vitro* the oocytes were stripped of cumulus cells and enucleated with a beveled micropipette at approximately 18 hours post maturation (hpm). Enucleation was confirmed in TL-HEPES medium plus bisbenzimidazole (Hoechst 33342, 3 µg/ml; Sigma). Individual donor cells were then placed into the perivitelline space of the recipient oocyte. The bovine oocyte cytoplasm and the donor nucleus (NT unit) are fused together using electrofusion techniques. One fusion pulse consisting of 90 V for 15 µsec was applied to the NT unit. This occurred at 24 hours post-initiation of

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1 maturation (hpm) of the oocytes. The NT units were placed in CR1aa medium  
2 until 28 hpm.

3 The procedure used to artificially activate oocytes has been described  
4 elsewhere. NT unit activation was at 28 hpm. A brief description of the  
5 activation procedure is as follows: NT units were exposed for four min to  
6 ionomycin (5  $\mu$ M; CalBiochem, La Jolla, CA) in TL-HEPES supplemented with  
7 1 mg/ml BSA and then washed for five min in TL-HEPES supplemented with  
8 30 mg/ml BSA. The NT units were then transferred into a microdrop of CR1aa  
9 culture medium containing 0.2 mM DMAP (Sigma) and cultured at 38.5°C 5%  
10 CO<sub>2</sub> for four to five hours. The NT units were washed and then placed in a  
11 CR1aa medium plus 10% FCS and 6 mg/ml BSA in four well plates containing  
12 a confluent feeder layer of mouse embryonic fibroblasts (described below). The  
13 NT units were cultured for three more days at 38.5°C and 5% CO<sub>2</sub>. The culture  
14 medium was changed every three days until day 12 after the time of activation.  
15 At this time NT units reaching the desired cell number, i.e., about 50 cell  
16 number, were mechanically removed from the zona and used to produce  
17 embryonic cell lines. A photograph of an NT unit obtained as described above  
18 is contained in Figure 1.

#### 19 Fibroblast feeder layer

20 Primary cultures of embryonic fibroblasts were obtained from 14-16 day  
21 old murine fetuses. After the head, liver, heart and alimentary tract were  
22 aseptically removed, the embryos were minced and incubated for 30 minutes at  
23 37°C in prewarmed trypsin EDTA solution (0.05% trypsin/0.02% EDTA;  
24 GIBCO, Grand Island, NY). Fibroblast cells were plated in tissue culture flasks  
25 and cultured in alpha-MEM medium (BioWhittaker, Walkersville, MD) supple-  
26 mented with 10% fetal calf serum (FCS) (Hyclone, Logen, UT), penicillin (100

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1 IU/ml) and streptomycin (50 µl/ml). Three to four days after passage, embryonic  
2 fibroblasts, in 35 x 10 Nunc culture dishes (Baxter Scientific, McGaw Park, IL),  
3 were irradiated. The irradiated fibroblasts were grown and maintained in a  
4 humidified atmosphere with 5% CO<sub>2</sub> in air at 37°C. The culture plates which  
5 had a uniform monolayer of cells were then used to culture embryonic cell lines.

6 Production of embryonic cell line.

7 NT unit cells obtained as described above were washed and plated directly  
8 onto irradiated feeder fibroblast cells. These cells included those of the inner  
9 portion of the NT unit. The cells were maintained in a growth medium  
10 consisting of alpha MEM supplemented with 10% FCS and 0.1 mM beta-  
11 mercaptoethanol (Sigma). Growth medium was exchanged every two to three  
12 days. The initial colony was observed by the second day of culture. The colony  
13 was propagated and exhibits a similar morphology to previously disclosed mouse  
14 embryonic stem (ES) cells. Individual cells within the colony are not well  
15 defined and the perimeter of the colony is refractile and smooth in appearance.  
16 The cell colony appears to have a slower cell doubling time than mouse ES cells.  
17 Also, unlike bovine and porcine derived ES cells, the colony does not have an  
18 epithelial appearance thus far. Figures 2 through 5 are photographs of ES-like  
19 cell colonies obtained as described, *supra*.

20 Production of Differentiated Human Cells

21 The human embryonic cells obtained are transferred to a differentiation  
22 medium and cultured until differentiated human cell types are obtained.

23 RESULTS

24 Table 1. Human cells as donor nuclei in NT unit production and development.

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**TABLE 1**

Cell type	No. NT units made	No. NT units 2 cell stage (%)	No. NT units to 4 - 16 cell stage (%)	No. NT units to 16 - 400 cell stage (%)
lymphocytes	18	12 (67%)	3 (17%)	0
oral cavity epithelium	34	18 (53%)	3 (9%)	1 (3%)
adult fibroblasts	46	4 (9%)	12 (4 cell; 26%) 8 (8-16 cells; 17.4%)	---

The one NT unit that developed a structure having greater than 16 cells was plated down onto a fibroblast feeder layer. This structure was attached to the feeder layer and started to propagate forming a colony with a ES cell-like morphology (*See, e.g., Figure 2*). Moreover, although the 4 to 16 cell stage structures were not used to try and produce an ES cell colony, it has been previously shown that this stage is capable of producing ES or ES-like cell lines (mouse, Eistetter et al., *Devel. Growth and Differ.*, 31:275-282 (1989); Bovine, Stice et al., 1996)). Therefore, it is expected that 4 - 16 cell stage NT units should also give rise to embryonic or stem-like cells and cell colonies.

Also, similar results were obtained upon fusion of an adult human keratinocyte cell line with an enucleated bovine oocyte, which was cultured in media comprising ACM, uridine, glucose, and 1000 IU of LIF. Out of 50 reconstructed embryos, 22 cleaved and one developed into a blastocyst at about day 12. This blastocyst was plated and the production of an ES cell line is ongoing.

While the present invention has been described and illustrated herein by reference to various specific materials, procedures, and examples, it is understood that the invention is not restricted to the particular material, combinations

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- 1 of materials, and procedures selected for that purpose. Numerous variations of such details can be implied and will be appreciated by those skilled in the art.



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WHAT IS CLAIMED IS:

1. A method of producing embryonic or stem-like cells comprising the following steps:

(i) inserting a desired differentiated human or mammalian cell or cell nucleus into an enucleated animal oocyte, wherein such oocyte is derived from a different animal species than the human or mammalian cell under conditions suitable for the formation of a nuclear transfer (NT) unit;

(ii) activating the resultant nuclear transfer unit;

(iii) culturing said activated nuclear transfer unit until greater than the 2-cell developmental stage; and

(iv) culturing cells obtained from said cultured NT units to obtain embryonic or stem-like cells.

2. The method of Claim 1, wherein the cell inserted into the enucleated animal oocyte is a human cell.

3. The method of Claim 2, wherein said human cell is an adult cell.

4. The method of Claim 2, wherein said human cell is an epithelial cell, keratinocyte, lymphocyte or fibroblast.

5. The method of Claim 2, wherein the oocytes are obtained from a mammal.

6. The method of Claim 5, wherein the animal oocyte is obtained

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from an ungulate.

7. The method of Claim 6, wherein said ungulate is selected from the group consisting of bovine, ovine, porcine, equine, caprine, and buffalo.
8. The method of Claim 1, wherein the enucleated oocyte is matured prior to enucleation.
9. The method of Claim 1, wherein the fused nuclear transfer units are activated *in vitro*.
10. The method of Claim 1, wherein the activated nuclear transfer units are cultured on a feeder layer culture.
11. The method of Claim 10, wherein the feeder layer comprises fibroblasts.
12. The method of Claim 1, wherein in step (iv) cells from a NT unit having 16 cells or more are cultured on a feeder cell layer.
13. The method of Claim 12, wherein said feeder cell layer comprises fibroblasts.
14. The method of Claim 13, wherein said fibroblasts comprise mouse embryonic fibroblasts.

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15. The method of Claim 1, wherein the resultant embryonic or stem-like cells are induced to differentiate.

16. The method of Claim 2, wherein the resultant embryonic or stem-like cells are induced to differentiate.

17. The method of Claim 1, wherein fusion is effected by electrofusion.

18. Embryonic or stem-like cells obtained according to the method of Claim 1.

19. Human embryonic or stem-like cells obtained according to the method of Claim 2.

20. Human embryonic or stem-like cells obtained according to the method of Claim 3.

21. Human embryonic or stem-like cells obtained according to the method of Claim 4.

22. Human embryonic or stem-like cells obtained according to the method of Claim 6.

23. Human embryonic or stem-like cells obtained according to the method of Claim 7.

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24. Differentiated human cells obtained by the method of Claim 16.
25. The differentiated human cells of Claim 24, which are selected from the group consisting of neural cells, hematopoietic cells, pancreatic cells, muscle cells, cartilage cells, urinary cells, liver cells, spleen cells, reproductive cells, skin cells, intestinal cells, and stomach cells.
26. A method of therapy which comprises administering to a patient in need of cell transplantation therapy isogenic differentiated human cells according to Claim 24.
27. The method of Claim 26, wherein said cell transplantation therapy is effected to treat a disease or condition selected from the group consisting of Parkinson's disease, Huntington's disease, Alzheimer's disease, ALS, spinal cord defects or injuries, multiple sclerosis, muscular dystrophy, cystic fibrosis, liver disease, diabetes, heart disease, cartilage defects or injuries, burns, foot ulcers, vascular disease, urinary tract disease, AIDS and cancer.
28. The method of Claim 26, wherein the differentiated human cells are hematopoietic cells or neural cells.
29. The method of Claim 26, wherein the therapy is for treatment of Parkinson's disease and the differentiated cells are neural cells.
30. The method of Claim 26, wherein the therapy is for the treatment of cancer and the differentiated cells are hematopoietic cells.

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31. The differentiated human cells of Claim 24, which contain and express an inserted gene.

32. The method of Claim 1, wherein a desired gene is inserted, removed or modified in said embryonic or stem-like cells.

33. The method of Claim 32, wherein the desired gene encodes a therapeutic enzyme, a growth factor or a cytokine.

34. The method of Claim 32, wherein said embryonic or stem-like cells are human embryonic or stem-like cells.

35. The method of Claim 32, wherein the desired gene is removed, modified or deleted by homologous recombination.

36. The method of Claim 1, wherein the donor cell is genetically modified to impair the development of at least one of endoderm, ectoderm and mesoderm.

37. The method of Claim 1, wherein the donor cell is genetically modified to increase differentiation efficiency.

38. The method of Claim 36, wherein the cultured nuclear transfer unit is cultured in a media containing at least one capsase inhibitor.

39. The method of Claim 1, wherein the donor cell expresses a

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detectable label that is indicative of the expression of a particular cyclin.

40. The method of Claim 36, wherein the donor cell has been modified to alter the expression of a gene selected from the group consisting of SRF, MESP-1, HNF-4, beta-1, integrin, MSD, GATA-6, GATA-4, RNA helicase A, and H beta 58.

41. The method of Claim 37, wherein said donor cell has been genetically modified to introduce a DNA that provides for expression of the Q7 and/or Q9 genes.

42. The method of Claim 41, wherein said gene or genes are operably linked to a regulatable promoter.

43. The method of Claim 1, wherein the donor cell has been genetically modified to inhibit apoptosis.

44. The method of Claim 43, wherein reduced apoptosis is provided by altering expression of one or more genes selected from the group consisting of Bad, Bok, BH3, Bik, Blk, Hrk, BNIP3, Gim<sub>L</sub>, Bid, EGL-1, Bcl-XL, Bcl-w, Mcl-1, A1, Nr-13, BHRF-1, LMW5-HL, ORF16, Ks-Bcl-2, E1B-19K, and CED-9.

45. The method of Claim 44, wherein at least one of said genes is operably linked to an inducible promoter.

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46. A mammalian somatic cell that expresses a DNA that encodes a detectable marker, the expression of which is linked to a particular cyclin.

47. The cell of Claim 46, wherein the cyclin is selected from the group consisting of cyclin D1, D2, D3, B1, B2, E, A and H.

48. The cell of Claim 46, wherein the detectable marker is a fluorescent polypeptide.

49. The cell of Claim 48, wherein said mammalian cell is selected from the group consisting of human, primate, rodent, ungulate, canine, and feline cells.

50. The cell of Claim 48, wherein said cell is a human, bovine or primate cell.

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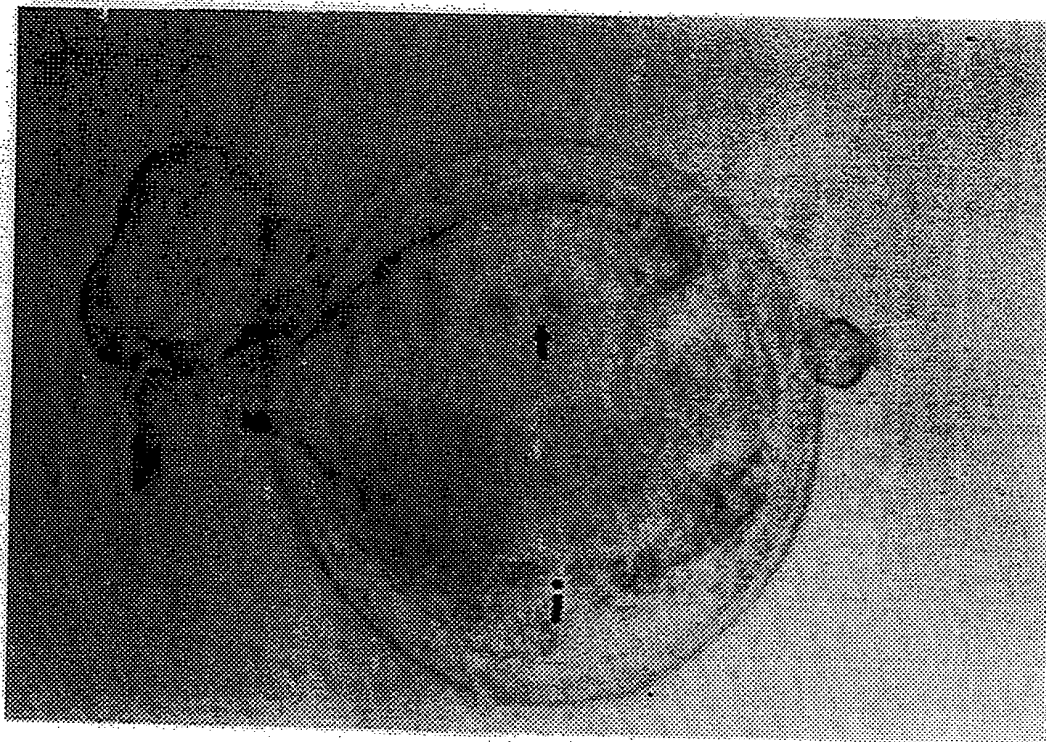


Fig. 1



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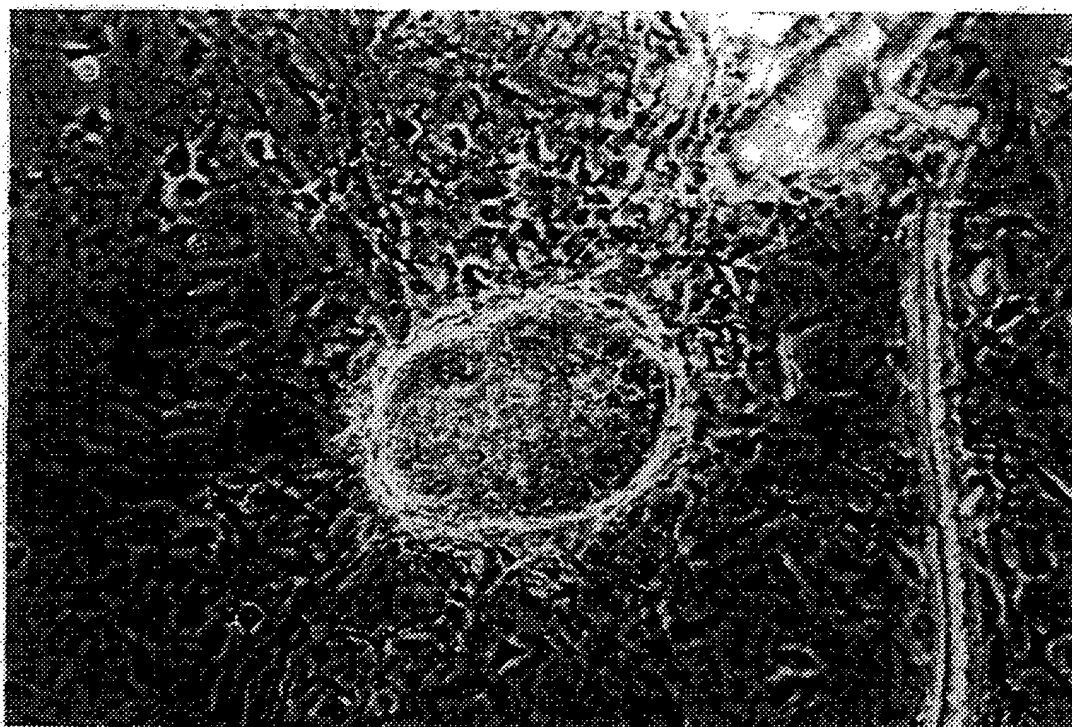


Fig. 2

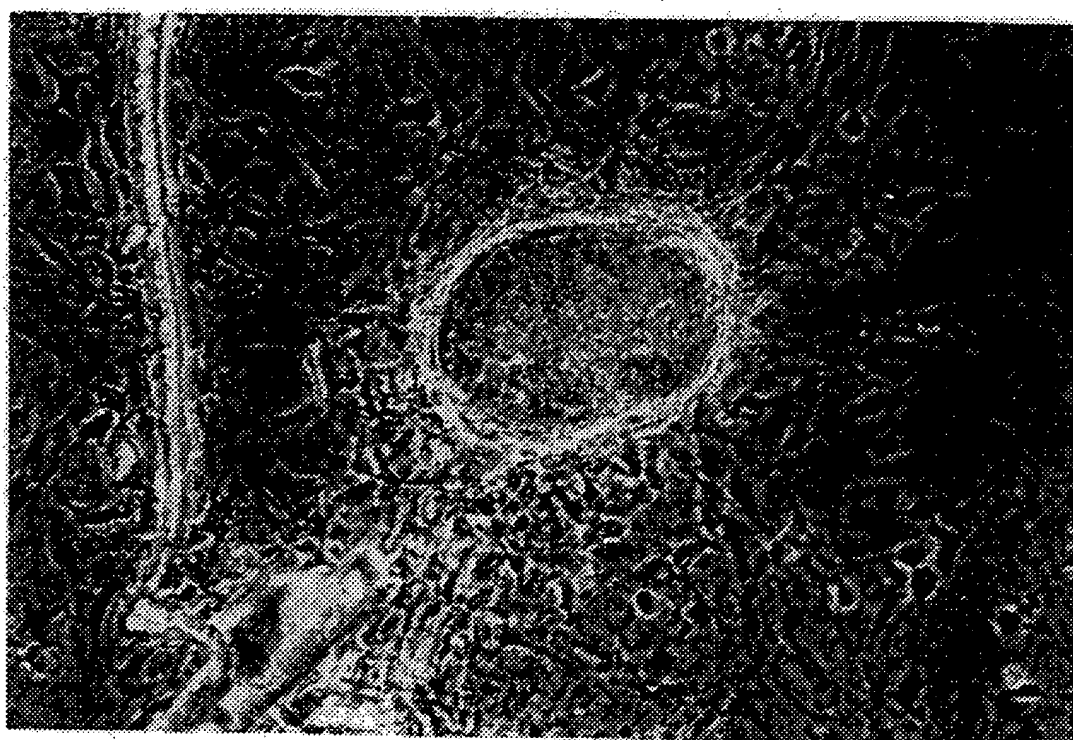


Fig. 3

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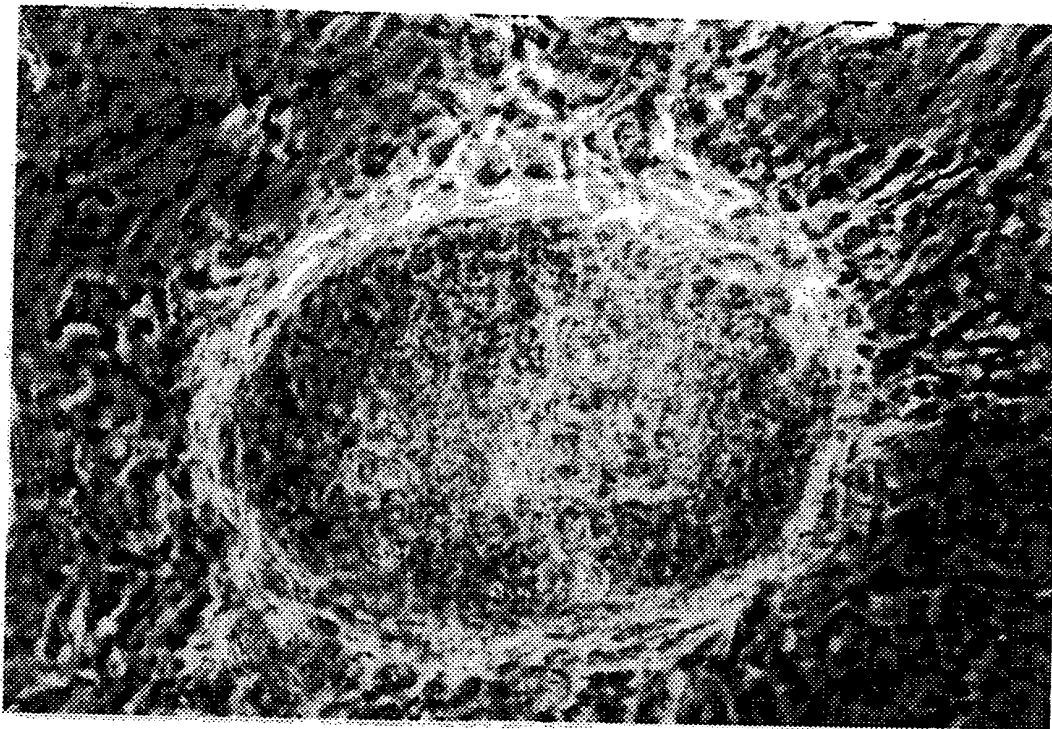


Fig. 4

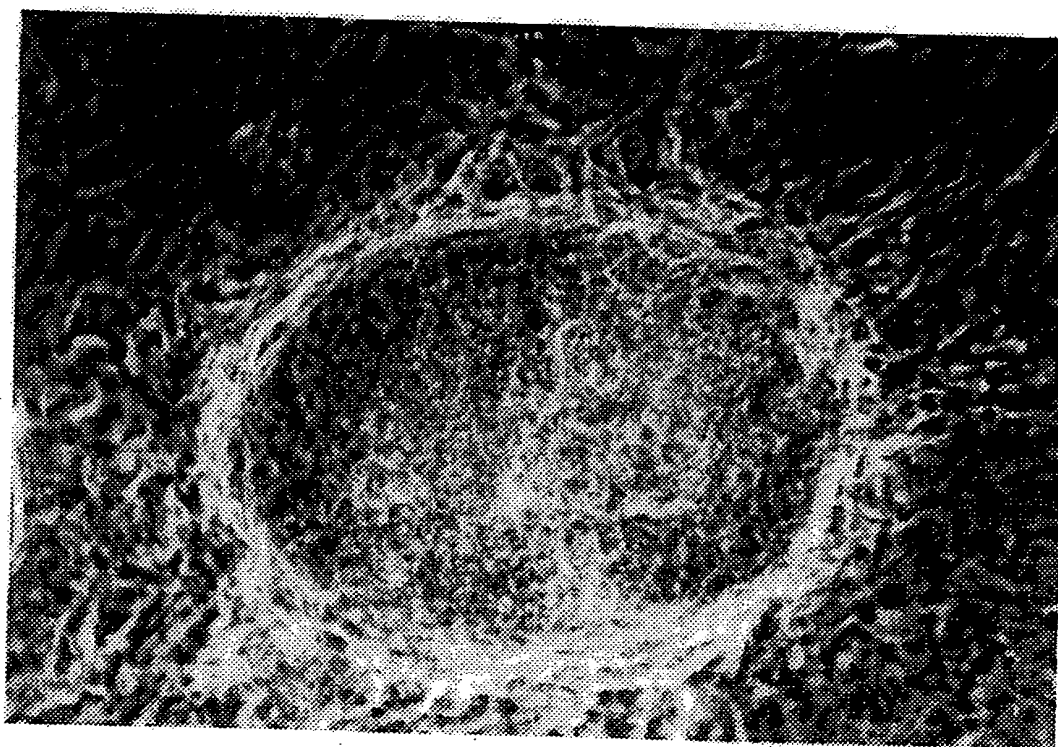


Fig. 5

## INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US99/04608

<b>A. CLASSIFICATION OF SUBJECT MATTER</b> IPC(6) : C12N 5/00, 15/00 US CL : 435/93.21, 325, 455; 800/13, 24 According to International Patent Classification (IPC) or to both national classification and IPC		
<b>B. FIELDS SEARCHED</b> Minimum documentation searched (classification system followed by classification symbols) U.S. : 435/93.21, 325, 455; 800/13, 24  Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched  Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) APS, MEDLINE, BIOSIS, EMBASE, CAPLUS search terms: embryonic stem cells, nuclear transplantation, cross-species nuclear transplantation		
<b>C. DOCUMENTS CONSIDERED TO BE RELEVANT</b>		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	WOLFE et al. Preimplantation Development of Embryos Produced by Intergeneric Nuclear Transplantation. Theriogenology. January 1990, Vol. 33, No. 1, page 350, see entire document.	1-25, 31-50
Y	COLLAS et al. Nuclear Transplantation by Microinjection of Inner Cell Mass and Granulosa Cell Nuclei. Molecular Reproduction and Development. 1994, Vol. 38, pages 264-267, see entire document.	1-25, 31-50
<input type="checkbox"/> Further documents are listed in the continuation of Box C. <input type="checkbox"/> See patent family annex.		
* "A"	Special categories of cited documents. document defining the general state of the art which is not considered to be of particular relevance	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
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"O"	document referring to an oral disclosure, use, exhibition or other means	
"P"	document published prior to the international filing date but later than the priority date claimed	"X" document member of the same patent family
Date of the actual completion of the international search 10 MAY 1999		Date of mailing of the international search report <b>14 JUN 1999</b>
Name and mailing address of the ISA US Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231 Facsimile No. (703) 305-3230		Authorized officer <i>R. Lawrence Fox</i> JILL D. MARTIN Telephone No. (703) 308-0196

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